

The activity of class I, II, III and IV alcohol dehydrogenase isoenzymes and aldehyde dehydrogenase in the sera of bladder cancer patients

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Objectives. Studies on alcohol dehydrogenase (ADH) and aldehyde dehydrogenase (ALDH) activity in the sera of patients with malignant neoplasms show that cancer cells in many organs may release ADH isoenzymes into the blood. The aim of this study was to investigate the differences in the activity of ADH isoenzymes and ALDH in the sera of patients with bladder cancer (BCa), and with different grades of the disease. **Material and Methods.** Blood samples were taken from 39 patients with BCa (15 patients with low-grade and 24 with high-grade BCa) and from 60 healthy subjects. Class III and IV of ADH and total ADH activity were measured using the photometric method, while class I and II ADH and ALDH activity using the fluorometric method with class-specific fluorogenic substrates. **Results.** The activity of the class I ADH isoenzyme and total ADH was significantly higher in the sera of BCa patients as compared to control group. Analysis of ALDH activity did not show statistically significant differences between the tested groups. Significantly higher total activity of ADH in comparison to control was found in both, low-grade and high-grade BCa group. The activity of ADH class I was also significantly higher in high-grade BCa group when compared to low-grade patients and controls. **Conclusion.** The increase of total ADH activity in the sera of BCa patients seems to be caused by isoenzymes released from cancerous cells. The higher activity of ADH I probably resulted from metastatic tumors as significant increase was detected only in the sera of high-grade bladder cancer patients.

Key words: alcohol dehydrogenase isoenzymes, aldehyde dehydrogenase, bladder cancer

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Abbreviations: ADH, alcohol dehydrogenase; ALDH, aldehyde dehydrogenase; BCa, bladder cancer; IARC, International Agency for Research on Cancer; NDMA, p-nitrosodimethylaniline; DNA, deoxyribonucleic acid; NAD, nicotinamide adenine dinucleotide; NADH, reduced form of nicotinamide adenine dinucleotide

INTRODUCTION

Alcohol consumption is the risk factor for over 5% of male and 1.7% of female cancer cases worldwide (Boffetta *et al.*, 2006). Despite the large number of published studies, questions about the definite role of ethanol drinking on bladder carcinogenesis remains without unequivocal answers. Components of alcoholic beverages and their metabolites, mostly acetaldehyde, are excreted

through the urinary tract and have been detected in urine, which can influence cancer initiation (Tsukamoto *et al.*, 1993).

Acetaldehyde is the product of ethanol oxidation catalyzed by alcohol dehydrogenase (ADH), and it is subsequently transformed into acetate in reaction with aldehyde dehydrogenase (ALDH). According to International Agency for Research on Cancer (IARC) ethanol and acetaldehyde belongs to group 1 of human carcinogens. Acetaldehyde is highly toxic and mutagenic substance, causing sister chromatid exchanges in human cells and interfering with DNA synthesis and repair at many sites in the genome, which results in tumor development (Li *et al.*, 2016; Amanuma *et al.*, 2015). Moreover, acetaldehyde, by binding to DNA and cellular proteins, forms adducts, which can activate proto-oncogenes, inactivate tumor suppressor genes in replicating cells and inhibit numerous important enzymes of DNA synthesis pathways (Balbo *et al.*, 2016).

In our recent study, we found significant increase of total ADH activity (more than 2.6 times) in bladder cancer cells, compared to healthy bladder tissue. The total activity of ALDH was lower in cancerous cells than in control group. Moreover, a significant increase of isoenzyme class III ADH was stated in cancer group as whole and in the high-grade bladder cancer (unpublished results). These findings suggest the participation of ADH in bladder cancer development by increased ability of cancer cells to metabolize ethanol into acetaldehyde, and also by causing glutathione depletion, generation of reactive oxygen species in consequence and finally induction of oxidative stress.

Studies on ADH and ALDH activity in the patients with malignant neoplasms showed that the cancer cells in many organs may release ADH isoenzymes to the blood, which is the reason of elevated activity of the specific isoenzymes in the sera of cancer patients (Orywal and Szmitkowski, 2016). The aim of this study was to investigate the differences in the activity of alcohol dehydrogenase isoenzymes and aldehyde dehydrogenase in the sera of patients with bladder cancer, and with different grades of bladder neoplasm. The results may supplement the knowledge about the disturbances of ADH and ALDH activity during the course of bladder cancer.

MATERIAL AND METHODS

Material

Blood samples were taken before surgery from 39 patients of University's Clinical Hospital in Białystok,

who was diagnosed with bladder cancer (31 males and 8 females; mean age 66 years, range 52–92 years). All patients were diagnosed with urothelial cell carcinoma. The patients were divided into two groups: 15 patients were classified as low-grade and 24 as high-grade bladder cancer. 19/39 patients were current smokers, 9 were ex-smokers and 11 were lifelong non-smokers. None of the patients had received chemotherapy, radiotherapy or immunotherapy before tissue collection. The control group was composed of blood samples taken from 60 healthy subjects (40 males and 20 females; mean age 58 years, range 51–68 years). 18/60 healthy volunteers were current smokers, 9 were ex-smokers and 33 were lifelong non-smokers. All of the patients from studied and control group had a history of only occasional alcohol consumption.

The research protocol was approved by the Medical University of Białystok's Human Care Committee located in Białystok, Poland (Approval No R-I-002/267/2015). All patients gave their informed consent for the examination.

Methods

Determination of total ADH activity. Total ADH activity was estimated using the photometric method with p-nitrosodimethylaniline (NDMA) as a substrate (Skursky *et al.*, 1979). The reaction mixture (2 mL) contained serum (0.1 mL), 1.8 mL of 26 μ M substrate solution in 0.1 M sodium phosphate buffer, pH 8.5 and 0.1 mL of mixture containing 0.25 M n-butanol and 5 mM NAD. The reduction of NDMA was monitored at 440 nm with Shimadzu UV/VIS 1202 spectrophotometer (Shimadzu Europa GmbH, Duisburg, Germany).

Determination of total ALDH activity. Aldehyde dehydrogenase activity was measured using the fluorogenic method based on the oxidation of 6-methoxy-2-naphthaldehyde to fluorescent 6-methoxy-2-naphthoate (Orywal *et al.*, 2011). The reaction mixture contained 60 μ L of serum, 60 μ L of substrate, 20 μ L of 11.4 mM NAD and 2.8 mL of 50 mM sodium phosphate buffer, pH 8.5. The mixture also contained 50 μ L of 12 mM solution of 4-methylpyrazole as a specific inhibitor of ADH activity. The fluorescence was read at the excitation wavelength of 310 and the emission wavelength of 360 nm with Shimadzu RF-5301 spectrofluorophotometer (Shimadzu Europa GmbH, Duisburg, Germany).

Determination of class I and II ADH isoenzymes. Class I and II ADH isoenzyme activity was measured using fluorogenic substrates (4-methoxy-1-naphthaldehyde for class I and 6-methoxy-2-naphthaldehyde for class II) in a reduction reaction according to Wierzychowski *et al.*, (Wierzychowski *et al.*, 1989). The assays were performed in a reaction mixture containing serum (60 μ L), substrate

(150 μ L of 300 μ M), NADH (100 μ L of 1 mM) and 0.1 M sodium phosphate buffer, pH 7.6 (2.69 mL). The measurements were performed with Shimadzu RF-5301 spectrofluorophotometer at the excitation wavelength of 316 nm for both substrates and the emission of 370 nm for class I and 360 nm for class II isoenzyme.

Determination of class III ADH isoenzyme. The assay mixture for class III alcohol dehydrogenase contained serum (100 μ L), formaldehyde as the substrate (100 μ L of 1 mM), glutathione (100 μ L of 1 mM) and NAD (240 μ L of 1.2 mM) in 0.1 M NaOH-pyrophosphate buffer, pH 8.0 (Jelski *et al.*, 2014). The final volume was 2 mL. The reduction of NAD was monitored at 340 nm and 25°C with Shimadzu UV/VIS 1202 spectrophotometer.

Determination of class IV ADH isoenzyme. The assay mixture for ADH class IV activity contained serum (50 μ L), m-nitrobenzaldehyde as the substrate (132 μ L of 80 μ M) and NADH (172 μ L of 86 μ M) in 0.1 M sodium phosphate buffer, pH 7.5 (Orywal *et al.*, 2013). The oxidation of NADH was monitored at 340 nm and 25°C with Shimadzu UV/VIS 1202 spectrophotometer.

Statistical analysis. The differences between tested and control groups were evaluated by Mann-Whitney U test. To test the hypothesis, that there is a difference between the two grades of bladder cancer, ANOVA rank Kruskal-Wallis test was performed. Data were presented as mean values \pm standard deviation and median values. Statistically significant differences were defined as comparisons resulting in $p < 0.05$.

RESULTS

The activity of total ADH, ALDH and ADH isoenzymes in the sera of bladder cancer patients was presented in Table 1. The total activity of alcohol dehydrogenase was significantly higher (about 8.3 times) in the sera of patients with bladder cancer than in healthy subjects ($p < 0.001$). The mean total activity of ADH was 7.90 IU/l in the bladder cancer group and 0.95 IU/l in the control group. The analysis of ALDH activity did not indicate significant differences between the tested group as whole and healthy individuals. The ratio of total ADH activity to ALDH activity in bladder cancer group was much higher (0.94) compared to the control group (0.13).

The comparison of ADH isoenzymes activities showed that the highest difference was exhibited by class I ADH. The mean activity of this class of isoenzymes in the cancer group increased more than 3 times (5.86 mIU/l) in comparison to the control level (1.89 mIU/l). The increase of ADH I activity was statistically significant ($p < 0.05$). The other tested classes of ADH

Table 1. The comparison of ADH isoenzymes and ALDH activity in the sera of bladder cancer patients.

Tested Group	ADH I Mean \pm S.D.	ADH II Mean \pm S.D.	ADH III Mean \pm S.D.	ADH IV Mean \pm S.D.	ADH Total Mean \pm S.D.	ALDH Total Mean \pm S.D.
Bladder cancer	5.86 \pm 7.56	16.80 \pm 10.79	15.91 \pm 15.75	14.67 \pm 16.22	7.90 \pm 7.74	8.42 \pm 7.39
Low-grade BCa	3.17 \pm 6.01	15.52 \pm 14.26	8.52 \pm 4.11	13.15 \pm 14.04	7.88 \pm 7.72	7.95 \pm 8.19
High-grade BCa	6.34 \pm 9.77	16.37 \pm 9.61	17.52 \pm 16.11	16.72 \pm 19.36	8.69 \pm 7.83	10.96 \pm 11.62
Control group	1.89 \pm 0.92	14.01 \pm 3.37	8.89 \pm 2.53	12.18 \pm 3.87	0.95 \pm 0.48	7.23 \pm 2.45
	$p^a=0.013$	$p^a=0.151$	$p^a=0.630$	$p^a=0.159$	$p^a<0.001$	$p^a=0.560$
	$p^b=0.098$	$p^b=0.099$	$p^b=0.608$	$p^b=0.729$	$p^b=0.049$	$p^b=0.667$
	$p^c=0.018$	$p^c=0.210$	$p^c=0.186$	$p^c=0.214$	$p^c<0.001$	$p^c=0.862$
	$p^d=0.201$	$p^d=0.201$	$p^d=0.200$	$p^d=0.577$	$p^d=0.694$	$p^d=0.768$

S.D., standard deviation. BCa, bladder cancer. Statistically significant differences were defined as comparisons resulting in $p < 0.05$. Data were expressed as mIU/l (ADH total – IU/l). p^a , cancer patients vs controls. p^b , low-grade BCa vs controls. p^c , high-grade BCa vs controls. p^d , low-grade BCa vs high-grade BCa

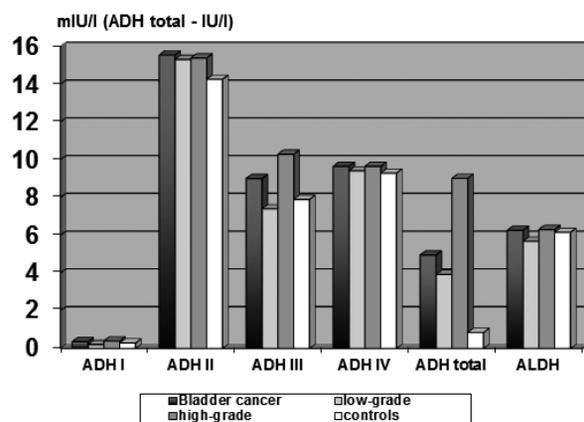


Figure 1. Comparison of ADH isoenzymes and ALDH activity in the sera of bladder cancer patients.

Data are expressed as median values of activity.

isoenzymes had higher activity in the sera of patients with bladder cancer but the differences were not statistically significant.

The comparison of ADH isoenzymes and ALDH activities in different grades of urinary bladder cancer was presented in the Fig. 1. Significantly higher ($p < 0.05$) total activity of ADH was found in the sera of both, low-grade (7.88 IU/l) and high-grade bladder cancer patients (8.69 IU/l), in comparison to healthy volunteers. The activity of ADH class I was significantly higher only in the high-grade bladder cancer group (6.34 mIU/l) compared to low-grade group (3.17 mIU/l) and controls. The other ADH isoenzymes did not exhibit any characteristic change of activity correlating with the bladder cancer grade. The total activity of ALDH also did not present significant differences between the different grades of the disease.

DISCUSSION

Urinary bladder cancer represents the 10th most common neoplasia worldwide, with the significant morbidity and mortality. This cancer is the second most prevalent malignant disease in elderly men owing to smoking and industrial exposures and the propensity of the urothelium for metastatic tumours (Jemal *et al.*, 2009). Bladder cancer has a variable metastatic potential and as the result most patients have more than one site of metastasis. Almost any organ can be subjected to bladder cancer metastasis but the most common sites are lymph nodes, bones, lungs, liver and peritoneum (Shinagare *et al.*, 2011). Clinically, 60% of bladder cancers are low-grade, whereas 25% of newly diagnosed bladder cancers are high-grade (Parkin *et al.*, 2000). Low-grade bladder cancer rarely invades the muscular wall of the bladder or spreads to the other organs. High-grade bladder cancer has a strong tendency to invade the muscular wall of the bladder and at least half of high-grade patients develop local and distant metastasis (Wu, 2005).

In our study, we found that the total activity of ADH was significantly higher in the sera of patients with bladder cancer compared to controls, however, the activity of ALDH was not different between the groups. Significantly higher total ADH activity was found in the sera of both, low-grade and high-grade bladder cancer patients. In our previous study, significant increase of ADH total activity was found in bladder cancer cells (low-grade and high-grade) in comparison to normal bladder tissue (un-

published results). These findings suggest increased ability to produce acetaldehyde and less capacity to remove it of the bladder cancer cells, compared to normal urinary bladder tissue, which can cause or intensify carcinogenesis in this organ. Our studies are consistent with the published results of ADH and ALDH activities in many malignant diseases. Significantly higher increase of total ADH activity was found both, in the cancer cells and in the sera of patients with neoplasms of esophagus, stomach, liver, pancreas, colorectal, kidney, endometrium and brain (Orywal & Szmitkowski, 2016). These studies confirmed that changes in enzyme activity in cancer tissue can be reflected in the sera of the patients probably due to releasing the enzyme during the malignancy course.

In the sera of bladder cancer patients we found significantly elevated activity of class I ADH isoenzyme. In most malignant diseases, patients have the same class of ADH isoenzymes concentration elevated in the serum as was in the cancer cells. In the oesophagus and stomach cancer, there was elevated activity of ADH IV, in pancreatic cancer – ADH III and in gynaecological cancers – ADH I, both in cancer tissue and in the sera of the patients (Orywal & Szmitkowski, 2016). During the course of renal cell carcinoma, elevated activity of ADH I was found in cancer cells and there was a tendency of ADH I activity to increase in the sera of patients in accordance with the advance of the disease. Significantly higher ADH class I activity was found in every stage (from II to IV) of kidney cancer compared to the control group (Orywal *et al.*, 2015). In the bladder cancer cells, we found significantly higher activity of ADH isoenzyme class III in the cancer group as whole, compared to healthy bladder cells (unpublished results). In sera of patients with bladder cancer, the activity of class III ADH was more than two times higher than in healthy individuals but the difference was not significant. However, increased activity of this class of ADH seemed to be the result of isoenzymes being released from bladder cancer cells.

It is interesting that in the sera of bladder cancer patients we found elevated levels of the other isoenzyme – ADH I, whose level in bladder cancer cells was similar to that in healthy tissue. It can be the result of isoenzyme being released from metastatic tumours e.g. liver cancer. Human liver contains almost all classes of alcohol dehydrogenase with the highest activity presented by class I. Chrostek and coworkers found that serum activity of class I ADH isoenzymes was significantly higher only in patients with metastatic liver tumors, while in primary tumors there was a tendency for this class activity to decrease (Chrostek & Szmitkowski, 2000). Metastatic cancers are infiltrating tumors and aggressively act on liver tissue leading to the release of the enzymes from normal liver cells. This hypothesis confirms the fact that the significant increase of the ADH I activity was found only in high-grade bladder cancer. These cancer types very often and easily develop distant metastasis. Furthermore, recent findings showed that polymorphism of alcohol dehydrogenase may be related to bladder cancer. Moderate drinkers with ADH I genotype ($\gamma 1\gamma 1$) appeared to have a threefold higher risk of bladder cancer compared to moderate drinkers with $\gamma 1\gamma 2$ and $\gamma 2\gamma 2$ genotype (Van Dijk *et al.*, 2001). Studies of Jelski and coworkers on breast cancer in women showed that despite of significantly lower ADH I activity in breast cancer cells, compared to normal parenchyma, its activity was higher in the sera of the patients (Jelski *et al.*, 2006a, Jelski *et al.*, 2006b). Significantly elevated activity of ADH I was

stated only in the blood of patients with stage IV breast cancer as the result of releasing isoenzymes from organs damaged by metastatic disease (Jelski *et al.*, 2006b).

In conclusion, we can state that the activity of class I ADH isoenzymes and the total activity of ADH were elevated in the sera of patients with bladder cancer as compared to the control group. The higher activity of ADH I was probably caused by metastatic tumors as significant increase was present detected only in the sera of high-grade bladder cancer patients. The increase of total ADH activity in the sera of bladder cancer patients can be a result of enzymatic disturbances in cancerous cells and isoenzymes (especially ADH III) being released from these cells.

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