

## ***Helicobacter pylori* increases expression of proapoptotic markers Fas and FasL on CD4 lymphocytes in children**

Aldona Kotłowska-Kmieć<sup>1</sup>✉, Alicja Bąkowska<sup>2</sup>, Adam Szarszewski<sup>3</sup>, Barbara Kamińska<sup>3</sup>, Grażyna Łuczak<sup>3</sup>, Wojciech Radys<sup>3</sup>, Piotr Landowski<sup>3</sup>, Jacek Brodzicki<sup>3</sup>, Maria Korzon<sup>3</sup> and Anna Liberek<sup>3</sup>

<sup>1</sup>Children's Hospital, Gdańsk, Poland; <sup>2</sup>Department of Immunopathology, <sup>3</sup>Chair and Department of Paediatrics, Paediatric Gastroenterology, Hepatology and Nutrition, Medical University of Gdańsk, Gdańsk, Poland

Received: 23 April, 2009; revised: 14 June, 2009; accepted: 22 June, 2009  
available on-line: 02 July, 2009

The pathomechanism of *Helicobacter pylori* action upon gastric mucosa and its role in the pathogenesis of gastritis have not been fully elucidated. The aim of this study was to evaluate the most prevalent lymphocyte subpopulations of the gastric mucosa in gastritis in children, as well as to evaluate the expression of Fas and Fas ligand receptors (FasL), periapoptotic markers of gastric mucosa lymphocytes before and after *H. pylori* eradication. Forty nine patients aged 6 to 17 years, investigated due to chronic abdominal pain, were studied. The obtained tissue samples were analysed by immunohistochemistry. Different lymphocyte subsets were quantified on the basis of surface antigen expression (CD3, CD4, CD8, CD20), secreted cytokines (IL-4, IL-6, IFN $\gamma$ ) and Fas and FasL proteins in the gastric mucosa. B and T helper lymphocytes were found to play a major role in the inflammatory infiltration in the gastric mucosa in children during *H. pylori* infection. Their expression was found to decrease after eradication. The enhanced expression of Fas receptor on lymphocytes before treatment and a decrease of this expression after eradication of *H. pylori* were shown. It was demonstrated that there is a correlation between CD4 and Fas receptor expression that may induce apoptosis of the helper lymphocytes in infected children.

**Keywords:** *H. pylori*, lymphocyte, T cell, apoptosis, Fas, FasL

### **INTRODUCTION**

Nowadays, it is accepted that as a result of *Helicobacter pylori* damaging action upon gastric mucosa the host's immunological mechanisms are activated (Dohil *et al.*, 1999). The lymphocyte subpopulation that plays the dominant role in inflammatory infiltration in children is still uncertain. Persistence of the inflammation, in spite of the induction of the host's immunological reactions, suggests that *H. pylori* has developed some mechanisms of evading immunosurveillance (Ernst & Gold, 1999; Wang *et al.*, 2001; Krauss- Etschmann *et al.*, 2005). One of them might be apoptosis. *H. pylori* induced apoptosis of

epithelial cells and lymphocytes appears to be enhanced by the expression of the so called "death receptors", including the Fas (CD95) and Fas ligand (CD95L) system (Souza *et al.*, 2006; Wisniewski *et al.*, 2008). A few recently published studies investigating paediatric population have shown an increase in the apoptosis index and proliferation of epithelium cells in *H. pylori* infected children as compared to healthy children. This suggests that eradication of *H. pylori* diminishes this process (Houghton *et al.*, 1999; Tytgat, 1992; Singh *et al.*, 2006). However, *H. pylori* might delay spontaneous lymphocyte apoptosis and prolong the survival of these cells or even inhibit T-cell proliferation (Schmees *et al.*, 2007). It can

✉Corresponding author: Aldona Kotłowska-Kmieć, Children's Hospital, Polanki 119, 80-308 Gdańsk, Poland; tel.: (48) 58 520 9315; fax: (48) 58 552 4741; e-mail: [akmiec@amg.gda.pl](mailto:akmiec@amg.gda.pl)

**Abbreviations:** Fas, receptor Fas; FasL, receptor Fas ligand; CD, cluster of differentiation antigen; IL, interleukin; INF, interferon; PAP, peroxidase-antiperoxidase.

also enhance the "fratricidal" death of lymphocytes by apoptosis, by concurrent increase in the expression of Fas and FasL on their surface (Sommer *et al.*, 1998; Houghton *et al.*, 1999; Koyama, 2000; Wang *et al.*, 2001). This may be another step towards solving the problem of eradication and elimination of the bacteria.

The aim of the study was to evaluate the prevalence of lymphocyte subpopulations and periaoptotic receptors Fas and FasL in the gastric mucosa during *H. pylori* infection and after eradication treatment in children. We also made an attempt to answer the question which of the lymphocyte subsets predominantly enter the apoptosis induction phase.

## PATIENTS AND METHODS

The study group consisted of 49 patients referred to the Endoscopy Unit of the Chair and Department of Paediatrics, Paediatric Gastroenterology, Hepatology and Nutrition of the Medical University of Gdańsk (Poland). They were examined because of chronic abdominal pain which was an indication for upper gastrointestinal endoscopy (gastroduodenofiberscope GIF 160-Olympus).

Children with a history of allergy, intake of antibiotics or anti-acid drugs within 30 days preceding the examination or with either primary or secondary malabsorption syndrome were excluded.

Forty nine patients aged 6 to 17 years (mean age  $12.86 \pm 2.59$ ) were examined. Thirty three of them were girls aged 6 to 17 years (mean age  $12.97 \pm 2.96$ ) and 16 of them were boys aged 9 to 15 years (mean age  $12.63 \pm 1.63$ ).

Patients were divided into three groups:

**Group I** — 12 children (7 girls and 5 boys) aged 7 to 17 years (mean age  $13.17 \pm 2.55$ ) without gastritis in histological examination and without *H. pylori* infection,

**Group II** — 10 children (5 girls and 5 boys) aged 10 to 17 years (mean age  $13.6 \pm 2.17$ ) with chronic gastritis and without *H. pylori* infection,

**Group III** — 27 children (21 girls and 6 boys) aged 6 to 17 years (mean age  $12.45 \pm 2.75$ ) with chronic gastritis and with *H. pylori* infection and,

**Group IIIA** — 11 children (6 girls and 5 boys) aged 9 to 17 years (mean age  $13 \pm 2.72$ ) from group III after eradication treatment.

Complete eradication was obtained in 5 children from this group. In the remaining 6 patients the bacteria were still detectable in spite of treatment.

In all patients included in this study upper gastrointestinal endoscopy, urease test (Clo-test) and histological examination of the gastric and duode-

nal biopsies were performed in order to assess the grade of gastritis and duodenitis according to the Sydney classification (Misiewicz *et al.*, 1994; Dixon *et al.*, 1996). All patients had macroscopic lesions of gastric and duodenal mucosa in endoscopic examination and were submitted to the routine therapy<sup>1</sup>. Mild and moderate gastritis was mainly observed in group I and II, while moderate and serious gastritis was found in group III.

Children with gastritis and co-existing *H. pylori* infection were subjected to triple therapy: omeprazole for 4 weeks (0.5 mg/kg per day), amoxicillin (40–50 mg/kg per day) and clarithromycin (15 mg/kg per day) for 7 days.

Four to eight weeks after the treatment was completed, the patients with *H. pylori* infection had a follow-up endoscopy in order to assess the eradication of *H. pylori* (according to the Polish *H. pylori* Infection Workgroup consensus, 1997)<sup>1</sup>. Samples from all patients were obtained for immunohistochemistry.

To confirm the *H. pylori* infection we performed the following tests: rapid urease test (Clo-test), Giemsa staining of the histological sections to visualise the bacteria under the optical microscope. The level of expression and localisation of *H. pylori* antigens in the gastric tissue was analysed by immunohistochemistry (immunoperoxidase and immunofluorescent method) with the use of specific antibodies (Ashton-Key *et al.*, 1996). A child was considered to be infected with *H. pylori* when a rapid urease test and immunohistochemistry were positive. A negative status was assumed when all tests gave concordant negative results.

**Immunohistochemistry.** One section of a sample was immediately frozen in liquid nitrogen and stored at  $-74^{\circ}\text{C}$ , the other one was routinely fixed in 4% phosphate-buffered formaldehyde solution and embedded in paraffin blocks according to standard procedure for immunohistochemistry and hematoxylin-eosin staining.

**Immunofluorescence method.** Frozen tissue sections (4  $\mu\text{m}$  thick) were incubated with polyclonal anti-*H. pylori* antibodies (Novocastra) (Ashton-Key *et al.*, 1996), monoclonal antibodies to CD4, Fas/CD95, FasL (Novocastra), CD8, CD20 (Dako) and anti-IL-4, IL-6, IFN $\gamma$  (Santa Cruz Biotechnology) for 16 h at  $4^{\circ}\text{C}$ . The sections were incubated for 30 min at room temperature with fluorescein-conjugated mouse, rabbit or goat antibodies (Novocastra, Santa Cruz Biotechnology and Dako, respectively), in a dark chamber. Negative controls for each immunohistochemical staining consisted of sections in which the primary antibody had been replaced by nonimmune serum and TBS (Tris-buffered saline).

<sup>1</sup>Management of *Helicobacter pylori* infection (2000 year). The consensus of Polish Working Group on *Helicobacter pylori* of the Polish Society for Gastroenterology (2001). *Pol Gastroenterol* 8: 11–18 (in Polish).

Sections were independently examined by two pathologists in four independent visual fields at 400× magnification. They were semiquantitatively scored as: 0 (negative), 1 (weakly positive), 2 (positive), and 3 (strongly positive). This scale was rearranged into a two point scale: 0 (negative) and 1 (positive).

**Immunoenzymatic-PAP method.** Paraffin sections of tissue samples (4 µm thick) were fixed on silan and prepared for immunohistochemical studies after antigen retrieval (*H. pylori* by incubation with trypsin, CD4, CD8, CD20, Fas and FasL by microwave procedure). Next, slides were blocked with: protein block serum (Dako, X-09090) for detection of *H. pylori*, IL-4, IL-6 and IFN $\gamma$ ; normal rabbit serum (Dako, 0902) for detection of CD4, CD8, CD20, Fas, and FasL. Finally, sections were incubated with antibodies to: *H. pylori* (Novocastra, NCL-HPp) (Ashton-Key *et al.*, 1996), CD4 (Novocastra, NCL-CD4), IL-4 (sc-6050), IL-6 (sc-1261), IFN $\gamma$  (sc-6050) (Santa Cruz Biotechnology) overnight at 4°C and with antibodies to: CD8 (M7103), CD20 (M07550 (Dako), Fas/CD95 (NCL-FAS-310), FasL (NCL-FAS-L) (Novocastra) 1 h at room temp. in a dark chamber. Antigen-antibody complexes were conjugated with peroxidase (*H. pylori*-peroxidase-conjugated rabbit immunoglobulin (Dako, P-217); IL-4, IL-6, IFN $\gamma$ -biotinylated anti-rabbit/mouse/goat antibodies and peroxidase-conjugated streptavidin (LSAB System, K-4368); CD4, CD8, CD20, Fas/CD95, FasL – rabbit anti-mouse antibody (Dako, Z-0259) and complex peroxidase-antiperoxidase of mouse serum (PAP Mouse, Dako, P-0850).

The reaction products were visualized with diaminobenzidine (DAB, Dako K-465).

The specificity of immunohistochemical staining was confirmed by negative and relevant positive controls included in each staining procedure. Sections were examined independently by two pathologists in four independent visual fields at 400× magnification and semiquantitatively scored as follows: lack of expression or very weak staining was considered as 0, focal expression – 1, >50% of the cells stained positive for the relevant antigen – 2, and >80% of the cells stained positive for the rele-

vant antigen – 3. This scale was rearranged into a two point scale as 0 (negative) and 1 (positive).

**Statistical analysis.** The results of the quantitative tests were expressed as mean  $\pm$  SEM. The frequency of different antigens was compared with Pearson chi-square and McNemara chi-square tests with Yate's correction. To test the correlation between variables we used nonparametric Spearman rank-order correlation analysis. Results were considered significant at a value of  $P < 0.05$ . Statistical analysis was performed using Statistica (data analysis software system), version 6.0, StatSoft Inc. (USA).

The Bioethics Committee of the Medical University of Gdańsk approved the study protocol.

## RESULTS

The assessment of the different lymphocyte subset participation in the gastric mucosa infiltrates was based on the percentage of patients, in whom the expression of various characteristic antigens or cytokines was found before and after the eradication. The results are presented in Table 1. Statistical analysis demonstrated that CD4 and CD20 antigens were significantly increased in group III (*H. pylori* infected patients) as compared to group IIIA (patients after eradication treatment).

In addition, there were a significantly higher number of CD20 antigens in group III in comparison to group I (patients without any signs of *H. pylori* infection and without microscopic inflammatory lesions in the gastric mucosa) and group II (patients with signs of gastritis). However, there were no significant differences between groups II and I.

We also analysed the localisation of lymphocytes within gastric mucosa. The data are shown in Table 2.

Analysis of periapoptotic markers of epithelium and lymphocytes after *H. pylori* infection is shown in Table 3.

The expression of Fas receptor was higher in *H. pylori* infected patients than in group I (patients

**Table 1. The percentage of patients with expression of antigens characteristic for studied lymphocyte subsets in inflammatory infiltrates in gastric mucosa.**

Number of children in studied groups n=49	Lymphocyte surface antigens (%)				Cytokines (%)		
	CD3	CD4	CD8	CD20	IL-4	IFN $\gamma$	IL-6
Group I n=12	100	90.9*	100	54.5*	91.7	66.7	83.3
Group II n=10	88.9	57.1*	100	70*	80	70	100
Group III n=27	100	100*	100	100*	80.8	69.2	96.3
Group IIIA n=11	100	81.8	100	81.8	81.8	54.5	100

\* $P < 0.05$

**Table 2. Analysis of different lymphocyte subset antigens in gastric mucosa.**

Lymphocyte surface antigens and cytokines	CD4 (%)	CD8 (%)	CD20 (%)	IL-4 (%)	INF $\gamma$ (%)	IL-6 (%)
Superficial localisation	16.3	83.7	14.3	75.5	32.7	93.9
Deep localisation	81.6	100	79.6	69.4	61.2	91.8

**Table 3. The percentage of patients with expression of selected periapoptotic markers in gastric mucosa.**

Number of children in studied groups n=43649	Periapoptotic markers (%)	
	Fas	FasL
Group I n=12	41.7 *	100
Group II n=10	33.3 *	100
Group III n=27	77.8 *	100
Group IIIA n=11	36.4	90

\* $P<0.05$ **Table 4. Analysis of the localisation of selected periapoptotic markers.**

Periapoptotic markers	Fas (%)	FasL (%)
Superficial localisation	6.25	97.3
Deep localisation	77.1	97.3

with normal histopathological examination result and without *H. pylori* infection). Similarly, the Fas receptor expression was significantly different in group III (*H. pylori* infected children) than in group II (patients with chronic gastritis, without *H. pylori* infection). Statistical analysis showed significantly increased expression of Fas receptor in group II as compared to group IIIA. After eradication treatment children from group IIIA had significantly decreased

expression of Fas receptor, which was comparable to the level in group I and II (Table 3).

Similarly to the lymphocyte subset localisation, we analysed the localisation of selected periapoptotic markers. The data are shown in Table 4.

The analysis of Fas and FasL receptor localisation showed that Fas receptor was localised in lamina propria in a high percentage of patients (Table 4).

In order to specify further which of the lymphocyte subsets may be the subject of enhanced apoptosis induction caused by *H. pylori* we analysed whether there was a correlation between examined lymphocytic subpopulations and Fas and FasL receptors. The data are shown in Table 5.

We found that in *H. pylori* infected patients, there is a positive correlation ( $R=0.436$ ,  $P<0.05$ ) between CD4 antigen expression, characteristic for T helper lymphocytes, and Fas receptor expression – one of the periapoptotic markers.

## DISCUSSION

The problem of *H. pylori* infection in paediatric population is still to be investigated (Ernst & Gold, 1999; Luzza *et al.*, 2000). Recently, a number of studies pointing towards Th1 lymphocytes as the dominant subpopulation in the gastric mucosa, both in adults and children were published (Suerbaum *et al.*, 2002; Luzza *et al.*, 2000; Maciorkowska *et al.*,

**Table 5. Analysis of correlation of lymphocyte subsets with Fas and FasL receptors in *H. pylori* infected patients.**

Antigens	Number of patients in group III n=27	$R_s$	$P$
CD 3 & Fas	23	0.128	0.56
CD 3 & FasL	14	0.112	0.70
CD 4 & Fas*	23	0.436	0.04*
CD 4 & FasL	14	0.018	0.95
CD 8 & Fas	24	0.202	0.34
CD 8 & FasL	15	0.059	0.83
CD 20 & Fas	23	0.099	0.65
CD 20 & FasL	15	0.313	0.26

\* $P<0.05$

1999; Sommer *et al.*, 1998). Our study demonstrates that CD8 T cytotoxic/suppressor lymphocyte antigens were present in almost all patients from the study population. IFN $\gamma$ , characteristic for Th1 lymphocytes, was present in more than 50% of patients; IL-4 and IL-6 secreted by Th2 lymphocytes were found in 80% and 90% of children, respectively. There were no statistically significant differences in secretion of those cytokines among the studied groups of patients. Similarly to CD3 and CD8 T lymphocytes, T helper lymphocytes (CD4) and B lymphocytes (CD20) were frequent in the inflammatory infiltrates of the gastric mucosa biopsies. We found that the B lymphocyte subset was significantly more common in *H. pylori* infected patients than in group I or group II (patients without *H. pylori* infection) (Maciorkowska *et al.*, 1999; Bussiere *et al.*, 2006). Interestingly, the percentage of CD20 mononuclear cells was markedly reduced in the studied population after eradication treatment. There was a statistically significant decrease in the percentage of CD4 lymphocytes in *H. pylori* infected patients after eradication as compared to the group of patients before treatment.

What is important, we confirmed the major contribution of Th lymphocyte (CD4) subpopulation in the inflammatory infiltration in the gastric mucosa of *H. pylori* infected children. It was, however, impossible to specify the lymphocyte subtype: Th1 or Th2.

There are two theories concerning *H. pylori* influence on apoptosis of lymphocytes in gastric mucosa. Studies based on the material obtained from adult patients indicate that *H. pylori* might delay spontaneous lymphocyte apoptosis and prolong survival of these cells. It results in the prolonged action of cytokines secreted by these cells and therefore augments their damaging action upon gastric mucosa (Moss *et al.*, 1996; Bussiere *et al.*, 2006). The findings of other authors suggest that *H. pylori* infection may accelerate the programmed death of immunocompetent cells or even inhibit proliferation of T-cells; hence they cannot eliminate the bacteria (Schmees *et al.*, 2007; Wang *et al.*, 2001; Houghton *et al.*, 1999).

New studies suggest that the presence of *H. pylori* stimulates apoptosis of gastric mucosal epithelial cells and lymphocytes *via* two different pathways: membrane involving so called "death receptors" (Fas and FasL) and mitochondrial involving cytochrome c/Apaf-1, Bcl-2 proteins (Bland *et al.*, 2006; Konturek *et al.*, 2003; Moss *et al.*, 1996). The expression of Fas receptor in healthy gastric mucosa is low (Houghton *et al.*, 1999; Konturek *et al.*, 2003). The enhanced expression of Fas antigen on epithelial cells and lymphocytes and of Fas ligand receptor on lymphocytes may be a result of the action of IFN $\gamma$ ,

which is a cytokine secreted mainly by Th1 helper lymphocyte subset (Moss *et al.*, 1996). In our study, we investigated the expression of Fas and FasL antigens and we found that Fas antigen expression on epithelial cells and gastric mucosal lymphocytes was the highest in *H. pylori* infected patients and significantly decreased after eradication. Our results are consistent with other published data, which highlight the crucial role of *H. pylori* antigen in Fas receptor expression on the cell surface and therefore emphasise the increased possibility of apoptosis stimulation depending on the expression of this receptor. Moreover, the possibility of programmed cell death reduction is present even after eradication treatment (Wang *et al.*, 2000). We found a very high expression of FasL receptor in all groups of investigated patients, slightly lower in patients after eradication treatment. This receptor is absent on lymphocytes under physiological conditions (Houghton *et al.*, 1999; Konturek *et al.*, 2003). Such a high expression of FasL receptors in our study population may be a result of lymphocytic infiltration, the presence of which was confirmed by immunohistochemistry in all studied groups. The interaction between so called "death receptors" (Fas, FasL) is a signal which may trigger apoptosis in cells expressing these receptors. According to some authors, the expression of FasL receptor in gastric mucosa is mainly related to *H. pylori* infection (Ishihara *et al.*, 2001; Souza *et al.*, 2006). Our results did not confirm these observations. We found that an increase in expressions of FasL receptor may also be caused by the inflammatory process in gastric mucosa without *H. pylori* infection.

To sum up, we found that in the studied paediatric population *H. pylori* infection significantly enhances the level of Fas receptor expression on the cell surface and thus augments the possibility of entering apoptosis induction phase by these cells.

To answer the question which of the lymphocyte subsets mainly undergoes apoptosis, we analysed the correlation between the differentiation antigens of various lymphocyte subsets with Fas and FasL receptors in the group of *H. pylori* infected children. There was a correlation between CD4 antigen of T helper cells and Fas receptor. This suggests that the gastric mucosa T lymphocytes are the subpopulation that may predominantly enter apoptosis induction phase during *H. pylori* infection in children. Similar results were reported by other authors (Wang *et al.*, 2000; 2001; Koyama, 2000).

## CONCLUSIONS

The increased apoptosis of T helper cells may be the cause of persistent infection without eliminating the bacteria from the patient's organism. The

presented results may confirm that *H. pylori* evades immunosurveillance by the programmed death of immunocompetent cells, especially CD4 helper lymphocytes.

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