

Intracellular mechanisms of tumor cells' immunoresistance

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One of the main mechanisms for avoiding immune response by cancer cells is mediated by inducing an immunosuppressive environment in the tumor following activation of immune checkpoints, i.e. PD-1 or CTLA-4 receptor inhibitors on T lymphocytes. Interaction inhibition between PD-1 or CTLA-4 and their ligands (PD-L1, CD80, and CD85) leads to unblocking of the T-lymphocyte function, and thus destroys cancer cells. Certain intracellular signaling pathways are also involved in the development of tumor cell immunoresistance. Immunosuppressive pathways' activation blocking may increase the immunological anti-tumor control.

Received: 08 October, 2019; revised: 25 February, 2020; accepted: 26 March, 2020; available on-line: 22 April, 2020

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Acknowledgements of Financial Support: This research project was funded by grant No. SUB/2/DN/19/001/2211, financed by the Medical University of Bialystok, Poland

Abbreviations: Akt, proteín kinase B; Bcl-xL, B-cell lymphoma-extra large transmembrane molecule; CTLA-4, cytotoxic T-lymphocyte-associated antigen 4; CY-202, seliciclib; dMMR, mismatch repair-deficient; IL-8, interleukin-8; LAG-3, lymphocyte activation gene 3; mCRC-metastatic colorectal cancer; MEK, activation of mitogen-activated protein kinase kinase; mTORC2, rapamycin-insensitive protein complex; NF-κB, nuclear factor κ-light-chain-enhancer of activated B cells; PD-L1, programmed death-ligand; PI3K, phosphoinositide 3-kinase; PTEN, phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase gene; TCR, T-cell receptor; TIM-3, mucin-domain-containing molecule-3; TIL, tumor-infiltrating lymphocytes; TME, tumor microenvironment; TNF-α, tumor necrosis factor α; Tregregulatory T cells; STAT3, activator of signal transducer and activator of transcription 3

INTRODUCTION

Over 8 million people die from cancer each year. According to epidemiological estimates, in the upcoming two decades, the number of deaths will increase by 2-fold among men, and by about 1/3 in the female population. This means that the scale of morbidity and, unfortunately, mortality due to colorectal carcinoma is continously on the rise. Undoubtedly, a significant contribution to the progression of this disease is the progress of civilization and the development of the world in which we live. Patients with colon cancer are a relatively heterogeneous group and thus it is difficult to design a single effective treatment pattern. A basic element of therapy is surgical treatment, which is aimed at obtaining intestinal tissue free of cancer. Chemotherapy complements surgical intervention and is mainly based on 5-fluorouracil/folic acid, capecitabine, oxaliplatin, or irinotecan. More and more often, classical chemotherapy is supplemented with a targeted biological therapy, involving administration of antibodies directed against specific molecular structures that stimulate cancer development. This therapy is intended for strictly selected (potentially sensitive) groups of patients and includes such drugs as bevacizumab, cetuximab, panitumumab, or regorafenib (https://www.nccn.org/). The continuing growth rate of colorectal carcinoma incidence indicates a necessity for intensified research, and forces to seek novel methods for early detection and more ideal treatment for patients suffering from this type of cancer. Therapy that is currently employed is undeniably unsatisfactory.

Recently discovered phenomenon of cancer cells avoiding the immune response has become the basis for the development of a new, groundbreaking direction in cancer treatment, i.e. immunooncology. The American Society for Clinical Oncology (ASCO) in their "Clinical Cancer Advances 2016" report, published in the Journal of Clinical Oncology, deemed immunotherapy to be the greatest advance in 2016. This breakthrough discovery was further acclaimed by awarding the 2018 Nobel Prize to James P Allison and Tasuku Honjo. Restoring proper immune system function has proven to be an effective strategy for fighting cancer. Thanks to the presence of antigens on the cancer cell surface, the immune system is able to recognize it and destroy it. Activation of the immune system cells occurs according to a two signal model. Interaction of the TCR (T-cell receptor) with the MHC molecule leads to antigen presentation and activation of specific T lymphocytes that achieve full effector functions only after ligand binding to the co-stimulatory molecules of lymphocytes. Lack of the second signal results in lymphocyte anergy and a deacrese in the immune system response. Apart from activating co-stimulatory molecules, there are also molecules with immunosuppressive activity, called the immune checkpoints, which include: PD-L1, CTLA-4, LAG-3, or TIM-3 (Lynch et al., 2016). The receptor pathway of programmed cell death 1 (PD-1) and its ligand PD-L1 are one of the most-studied immune checkpoints. PD-L1 is a transmembrane glycoprotein that is responsible for maintaining peripheral tolerance by limiting the T-lymphocyte activity, proliferation and effector functions. Results of large clinical trials clearly indicate that high PD-L1 expression directly correlates with cancer stage, metastasis and worse prognosis (Zhuan-Sun et al., 2016, Zhang et al., 2017; Wang et al., 2017). This protein is a valuable prognostic biomarker and a reliable indicator of treatment effectiveness for some types of cancer (Hamanishi et al., 2007; Xiang et al., 2017; Zhang et al., 2017).

PD-1/PD-L1 SIGNALING PATHWAY – A LINK INHIBITING THE ANTI-TUMOR IMMUNE RESPONSE

PD-1/PD-L1 signaling pathway is essential under physiological conditions, where it is responsible for

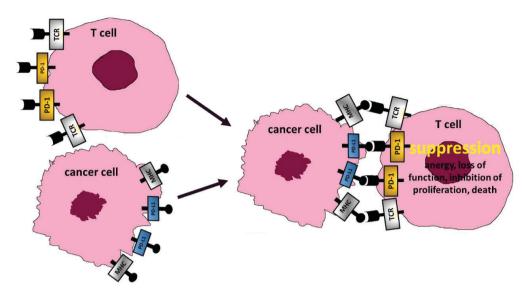


Figure 1. The PD-1/PD-L pathway.

MHC, major histocompatibility complex; PD-1, programmed death receptor 1; PD-L1, programmed death-ligand; TCR, T-cell receptor.

maintaining tolerance to its own antigens and prevents autoimmune disease development, while it also plays a key role in cancer cell "escape" from the immune surveillance (Fig. 1) (Nishimura et al., 2001, Boussiotis, 2016, Valentini et al., 2018). First described by Ishida and coworkers (Ishida et al., 1992) the PD-1 receptor is a glycoprotein receptor composed of 288 amino acids, encoded by the PDCD-1 gene located on chromosome 2 (Ishida et al., 1992; Zhang et al., 2004; Własiuk et al., 2016). It is continuously expressed, predominantly on activated T lymphocytes and macrophages, while the presence of this receptor on B lymphocyte and NK cell surfaces requires induction (Valentini et al., 2018). NF-ATc1, nuclear factor c1 of activated T lymphocytes, plays an important role in regulation of PD-1 expression. Its blocking reduces PD-1 expression while a mutation in the coding gene results in a complete lack of PD-1 expression (Oestreich et al., 2008). The primary role of the PD-1 receptor is to inhibit the T lymphocyte function, which is observed after receptor binding to one of the ligands on APC cells. This leads to slower cell metabolism, and thus lymphocyte depletion in their effector functions. Production of TNF-γ, TNF-α, and IL-2 is inhibited, and the amount of the anti-apoptotic Bcl-xL protein decreases, promoting the apoptosis process. While the CTLA-4 receptors inhibit T cell activation, mainly on the basis of competing for molecule CD80/CD86, the PD-1 receptor and its ligands constitute a separate signaling pathway (Riella et al., 2012). Two ligands for the PD-1 receptor, PD-L1 (B7-H1, CD274), programmed death molecule 1, and PD-L2 (B7-DC, ĈD273), programmed death molecule 2, were identified. They are transmembrane type I glycoproteins with IgV- and IgC-like domains. PD-L1 is expressed on the surface of the T and B lymphocytes, macrophages, dendritic cells, and numerous non-hematopoietic cells, whereas PD-L2 expression is more limited (Grzywnowicz & Giannopoulos, 2012). PD-L1 expression was also detected on the cell surfaces of numerous types of cancer, including bladder, kidney, ovarian, lung, and melanoma, which was associated with poorer immune response and worse prognosis

for patients. The large amounts of PD-L1 help some cancer cells to hide from an immune attack (Feld et al., 2016; Lee et al., 2016; Maleki et al., 2017; Xiang et al., 2017; Rom-Jurek et al., 2018). The presence of this protein has been also demonstrated in colorectal carcinoma cells. Valentini et al. confirmed its expression on both, the tumor cells and tumor-infiltrating lymphocytes (TIL) (Valentini et al., 2018). Although preliminary trials did not suggest a significant role of immunotherapy in the treatment of colorectal carcinoma (CRC), Le and coworkers (Le et al., 2015) showed a marked increase in an objective immune response rate and progression-free survival rate in patients treated with pembrolizumab. Results published in 2017 and 2018 established the PD-1 inhibitors, pembrolizumab and nivolumab, and the combination of nivolumab with the CTLA-4 inhibitor ipilimumab, as effective options for mismatch repair-deficient (dMMR) metastatic colorectal cancer (mCRC) (Le et al., 2015; Smith et al., 2018). Immunotherapeutic agents have grown in popularity for treating mismatch repair-deficient metastatic colorectal cancer (mCRC), becoming the standard of care in the second line. In turn, preliminary analyses of early stages of current research in patients with metastatic colorectal cancer demonstrated promising atezolizumab activity (anti-PD-L1 antibody) when used together with chemotherapy and/or targeted therapy with cobimetinib, the MEK inhibitor (Tapia et al., 2018). Despite initial enthusiasm, the IMblaze370 trial failed to improve survival over standard third-line therapy for patients with chemorefractory metastatic colorectal cancer and microsatellite-stable disease (Eng et al., 2019). However, recent results from phase II of the CCTG CO.26 study presented at the 2019 American Society of Clinical Oncology Annual Meeting clearly showed that the combination of the anti-PD-L1 antibody, durvalumab, and the anti-CTLA-4 antibody, tremelimumab, extended the median overall survival (OS) by 2.5 months when compared with the best supportive care alone in patients with microsatellite stable, refractory advanced CRC (6.6 vs 4.1 months; HR, 0.72; 95% CI, 0.54–0.97; P=0.07) (Chen et al., 2019).

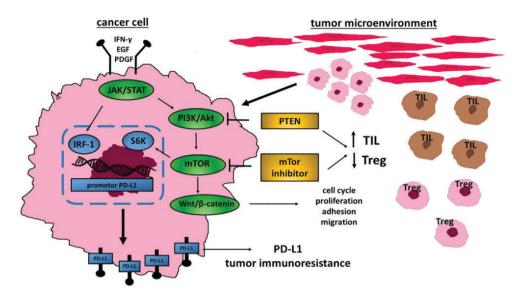


Figure 2. Selected intracellular mechanisms of tumor cell immunoresistance.

Akt, protein kinase B; IRF-1, interferon regulatory factor 6; JAK, Janus kinase; mTOR, mammalian target of rapamycin kinase; PD-L1, programmed death-ligand; Pl3K, phosphoinositide 3-kinase; S6K, ribosomal protein S6 kinase; PTEN, phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase gene; STAT, activator of transcription protein; TIL, tumor-infiltrating lymphocytes; Treg, regulatory T cell; WNT/b, catenin signal transduction pathways made up of proteins that pass signals into a cell through cell surface receptors.

The immune checkpoint blockade therapy has achieved a remarkable success in the treatment of patients with various cancer types and is a promising novel tool in oncology. Yet, only a limited fraction of patients derive a clinical gain. The results of large clinical trials show how, despite the exceptional survival benefit obtained with immune checkpoint blockade, some populations of patients (40-60%) will not avail from this therapy (Maleki et al., 2017; Catalano et al., 2019). Unfortunately, factors that influence the therapy response remain not fully known. Detection of PD-L1 on the surface of tumor cells is an important and so far the most common clinically detected biomarker for predicting patient response to the anti-PD-1/PD-L1 therapy (Herbst et al., 2014; Topalian et al., 2014; Ansell et al., 2015; Le et al., 2015; Maleki et al., 2017). There are also some interesting data indicatung the tumor microenvironment, the indole 2,3-dioxygenase, and finally mutational landscape, as biomarkers for predicting the response to immunotherapy (Ji et al., 2012; Holmgaard et al., 2013; Anagnostou et al., 2017; Maleki et al., 2017). It has been demonstrated that tumors with many somatic mutations due to mismatch-repair defects are more susceptible to the immune checkpoint blockade than tumors lacking this property (Le et al., 2015; Deppert & Bruns, 2016). Thus, immunogenicity of the tumor antigen T-cell epitopes may also be significant for treatment success or failure. Taking into account the dynamic nature of the immune system and the multiple factors involved in the complex antitumor immune response, developing biomarkers for immunotherapeutics can be quite a challenge.

It is worth noting that using immunotherapy is costly and might have some serious adverse events associated with excessive immune activation, termed immune-related adverse events (Bajwa et al., 2019). This upregulation can potentially affect one or more organs, leading to pneumonitis, hypothyroidism, hepatitis, colitis, as well as more general adverse events, such as fatigue, diarrhea, and rash (Ai & Curran, 2015; Turnis et al., 2015; Bajwa et al., 2019; Da1 et al., 2020).

Many events can be life-threatening, which requires discontinuation of treatment or long-term corticosteroids (Bertrand *et al.*, 2015).

INTRACELLULAR MECHANISMS OF TUMOR CELL IMMUNE-RESISTANCE

The PI3K/Akt pathway plays a key role in regulation of processes related to cell growth, metabolism, survival and proliferation. Akt, a serine/threonine protein kinase, also called protein kinase B, is the main signal transducer in this pathway. Increased expression of this kinase is observed in numerous cancers, including breast, lung, and prostate cancer (Lastwika et al., 2016). Its main cause is a mutation in the genes encoding Akt isoforms or amplification and activating gene mutation of the catalytic subunit of PI3K (the PIK3CA mutation). Increased Akt kinase activity is not usually a sufficient factor responsible for initiating the oncogenesis process, but it contributes to tumor progression by inhibiting apoptosis, promoting proliferation, migration and invasion. Moreover, recent studies indicate that the PI3K/ Akt/mTOR pathway is also involved in the development of tumor cell immune-resistance. Lastwikaand coworkers (Lastwika et al., 2016) showed that the m embrane PD-L1 expression in human lung cancer cells is significantly associated with mTOR kinase activation. It was proven that oncogenic activation of the Akt/mTOR pathway promotes the immune escape through enhanced PD-L1 expression (Fig. 2). Still, use of mTOR inhibitor in combination with the PD-1 antibody in the mouse lung cancer model is the cause of a significant inhibition of tumor growth, increase in TIL number, and a significant reduction in Treg number. These results indicate a significant contribution of the PI3K/Akt/mTOR pathway not only in the regulation of PD-L1 expression, but also in the development of an immunosuppressive tumor microenvironment (TME) (Lastwika et al., 2016). The data in the literature suggest the existence of two mechanisms responsible for the induction of PD-L1 expression in tumor cells. The first one is related to the reaction to

changes occurring in the tumor microenvironment. Increased PD-L1 expression is a response to TIL lymphocyte attack, the presence of which indicates immunogenicity of a given tumor, and whose efficacy is significantly reduced in the presence of PD-L1 (Parsa et al., 2007). Tumor-infiltrating lymphocytes (TIL) are a key component of the TME, reflecting the host antitumor immune response, defined as lymphocytes that surround, attack and eradicate tumor cells (Horton & Gajewski, 2018). TILs are thought to be suppressed by multiple immune inhibitory molecules in the tumor microenvironment and this suppression has been associated with tumor progression. Sobral-Leite et al. showed a clear association between TILs and PIK3CA mutations. Assessment of TIL subsets in terms of PI3K changes indicated that tumors with a PIK3CA mutation often possess more CD8+ cells (subset of TILs) (Sobral-Leite et al., 2019). Enhancement of CD8+T cell infiltration within the tumor tissue has been also observed after treatment with PI3K pathway inhibitors (selective PI3K β (p110 β), isoform inhibitor GSK2636771, pan-PI3K inhibitor LY29002). Increase in the number of CD8+T cell caused tumor burden and significant survival benefits in various animal models of cancer (O'Donnell et al., 2017). Reduction in the CD8+ numbers and high numbers of Treg in the tumor infiltrate are associated with a poor prognosis (Zeng, 2017). Tregs represent a unique CD4+ T-cell subpopulation that induces immunological self tolerance and are in charge of suppressing activation and proliferation of autoreactive lymphocytes. Recently, they have been shown to migrate into tumors and suppress an effective anticancer response in the tumor microenvironment (Beyer & Schultze, 2009; Onishi et al., 2014). Their depletion is effective in enhancing immune responses. Abu-Eid et al. showed that PI3K-Akt inhibitors caused a significant antitumor effect which was Treg dependent. They inhibited activation and proliferation of Tregs with a minimal effect on conventional T cells. This effect has been observed both in vitro (human and murine CD4 T cells) and in vivo, in naive and tumor-bearing mice (Abu-Eid et al., 2014).

The second mechanism responsible forinduction of PD-L1 expression, independent of the immune system activity, assumes a PTEN tumor suppressor gene mutation. This leads to loss of ability to inhibit the PI3K/ Akt/mTOR/S6K pathway, which consequently results in an increased PD-L1 expression (Fig. 2; Parsa et al., 2007). The phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase gene (PTEN) often mutates in the case of glioblastoma, melanoma, gastric, ovarian, kidney, breast, and lung cancers (Crowell et al., 2007). Moreover, activation of mTORC2 (rapamycin-insensitive protein complex) interferes with PTEN-loss Treg stability and their ability to differentiate (Huynh et al., 2015). De la Iglesia and others (De la Iglesia et al., 2008) demonstrated that the PTENloss glioblastoma do not show transcriptional repression of the interleukin-8 gene due to lack of binding of transcription factors (activator of signal transducer and activator of transcription 3-STAT3) to the IL-8 promoter. This resulted in increased transcription and expression of IL-8, which promotes glioblastoma cell proliferation and invasiveness only in a genetic PTEN-loss context (De la Iglesia et al., 2008). This relationship between PTEN-loss and a selective upregulation of IL-8 signaling has been also demonstrated in prostate carcinoma (Maxwell et al., 2013). Over-activation of the PI3K-AKT-mTOR pathway directly affects TME through upregulation of IL-8 (Conciatori et al., 2018). Furthermore, elevations of serum IL-8 concentrations reflect tumour growth dynamics

and have been considered as biomarkers for predicting response to the anti-PD-1 therapy in non-small cell lung cancer and melanoma. The most recent data from clinical and preclinical studies indicate that PI3K/Akt/mTOR inhibition may have a double benefit. On one hand, it may limit tumor growth by inhibiting cell proliferation, migration, and survival. On another hand, it may increase the immunological anti-tumor control by blocking activation of the immunosuppressive pathways and strengthening the internal immunity mechanisms (Xue et al., 2015).

Another pathway involved in the phenomenon of immunosuppression, or cancer cell "escape" from the immune system surveillance, is the intracellular JAK/ STAT. JAK and STAT (mainly STAT3 and STAT5) proteins control many processes associated with tumor development, including cell division and viability, new blood vessel formation, as well as immunoresistance. Increased activity of these proteins is observed in many cancers. Marzec et al. showed that the amplifiable 9p24 region contains the JAK2 gene, as well as the gene encoding PD-L1, which contributes to the development of Hodgkin's lymphoma due to weakening of the anti-tumor immune response (Marzec et al., 2008). Moreover, in patients with colorectal cancer and melanoma, who are carriers of the inactivating JAK1/2 gene mutation, a complete lack of PD-L1 protein expression is observed, resulting in resistance to therapy with inhibitors of immune checkpoints (anti-PD-1) (Shin et al., 2017). Another important factor regulating PD-L1 expression is the STAT3 protein. It was shown that in patients with lymphoma, characterized by NPM-ALK presence, increased activity of this protein leads to the PD-L1 over-expression (Green et al., 2010; Lastwika et al., 2016). NPM-ALK fusion was also shown to have the ability to activate STAT3, which is an indispensable factor for stimulating gene transcription of PD-L1 (Fig. 2; Davis et al., 2010). It is worth emphasizing that the STAT3 protein negatively regulates the immune response. It inhibits the Th1 activity by increasing expression of certain immunosuppressive factors, including IL-10 (Williams et al., 2004). It was also noticed that myeloid cells displaying the active form of the STAT3 protein inhibit the anti-tumor immune response (Yu et al., 2007). Research conducted by Wang et al. showed that blocking the STAT3 protein in B16 murine melanoma tumors results in a tumor mass reduction and increased T lymphocyte infiltration (Wang et al., 2004).

Looking for new therapeutic strategies combining anti-neoplastic activity with activation of the anti-tumor immune response is an especially urgent task for researchers, doctors, and the pharmaceutical industry. Growing knowledge in the field of molecular biology and cancer genetics has contributed to the discovery of new, effective therapeutic methods based on the intracellular signal transmission. Introduction of monoclonal antibodies and tyrosine kinase inhibitors for wide use was undoubtedly a huge success and a significant step in the fight against cancer. The special role of proteins that are overexpressed as a result of gene mutation or amplification in the course of the neoplastic process was recognized. High hopes are associated with micromolecule drugs with the ability to inhibit EGFR, VEGFR, intracellular signaling pathways, such as Ras/Raf/MEK/ERK, PI3K/Akt/mTOR, as well as cyclin-dependent kinase inhibitors, such as roscovitine, for example. Roscovitine, 2-[[9-(1-methylethyl)-6-[(phenylmethyl)amino]-9H-purin-2-yl[amino]-1-butanol also called CY-202 or seliciclib, is a low molecular weight purine derivative with a characteristic ring structure. It belongs to the cyclin-dependent kinase (CDK) protein inhibitors that play a key role in regulating the cell cycle, promoting its progression or transition between the individual phases. Increased CDK expression is observed in tumor cells, which may be the cause of cell cycle deregulation. CDK inhibitors inhibit cell division and induce programmed cell death. Roscovitine has the ability to inhibit numerous cyclin-dependent kinases, including: CDK1, CDK2, CDK5, CDK7, and CDK9, but also other kinases, such as CaMK2, CK1α, CK1δ, DYRK1A, EPHB2, ERK1, ERK2, FAK, and IRAK4. This compound also blocks the RNA II polymerase, lowers expression of Bcl-2, Mcl-1, and XIAP genes, and increases p53 expression, which in turn leads to cell death via apoptosis. The results of the latest research indicate that by inhibiting Cdk5 and blocking p53 degradation, roscovitine significantly weakens PD-L1 expression, promoting the anti-tumor immune response (Cortez et al., 2015; Dorand et al., 2016).

Despite considerable progress in treating cancer in recent years, designing an efficient anti-neoplastic therapy is still a serious challenge for contemporary oncology. Lack of sufficient knowledge on the tumorigenesis process, the mechanisms of drug resistance, or finally the escape of cancer cells from the immune system is the cause of the uneven and difficult fight against cancer.

REFERENCES

- Abu-Eid R, Samara RN, Ozbun L, Abdalla MY, Berzofsky JA, Friedman KM, Mkrtichyan M, Khleif SN (2014) Selective inhibition of regulatory T cells by targeting the PI3K-Akt pathway. *Cancer Immunol. Res.* 2: 1080–1089. https://doi.org/10.1158/2326-6066.CIR-14-0095
- Ai M, Curran MA (2015) Immune checkpoint combinations from mouse to man. Cancer Immunol. Immunother. 64: 885–892. https://doi. org/10.1007/s00262-014-1650-8
- Anagnostou V, Smith KN, Forde PM, Niknafs N, Bhattacharya R, White J, Zhang T, Adleff V, Phallen J, Wali N, Hruban C, Guthrie VB, Rodgers K, Naidoo J, Kang H, Sharfman W, Georgiades C, Verde F, Illei P, Li QK, Gabrielson E, Brock MV, Zahnow CA, Baylin SB, Scharpf RB, Brahmer JR, Karchin R, Pardoll DM, Velculescu VE (2017) Evolution of neoantigen landscape during immune checkpoint blockade in non-small cell lung cancer. Cancer Discap. 7: 264–276. https://doi.org/10.1158/2159-8290.CD-16-0828
- immune checkpoint blockaue in non-sinal cen lung cancer. Camer Discor. 7: 264–276. https://doi.org/10.1158/2159-8290.CD-16-0828
 Ansell SM, Lesokhin AM, Borrello I, Halwani A, Scott EC, Gutierrez M, Schuster SJ, Millenson MM, Cattry D, Freeman GJ, Rodig SJ, Chapuy B, Ligon AH, Zhu L, Grosso JF, Kim SY, Timmerman JM, Shipp MA, Armand P (2015) PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. N. Engl. J. Med. 372: 311–319. https://doi.org/10.1056/NEJMoa1411087
- Bajwa R, Cheema A, Khan T (2019) Adverse effects of immune check-point inhibitors (programmed death-1 inhibitors and cytotoxic T-lymphocyte-associated protein-4 inhibitors): results of a retrospective study. J. Clin. Med. Res. 11: 225–236. https://doi.org/10.14740/jocmr3750
- Bertrand A, Kostine M, Barnetche T, Truchetet ME, Schaeverbeke T (2015) Immune related adverse events associated with anti-CTLA-4 antibodies: systematic review and meta-analysis. *BMC Med.* **13**: 211. https://doi.org/10.1186/s12916-015-0455-8
- Beyer M, Schultze JL (2009) Regulatory T cells: major players in the tumor microenvironment. Curr. Pharm. Des. 15: 1879–1892. https:// doi.org/10.2174/138161209788453211
- Boussiotis VA (2016) Molecular and biochemical aspects of the PD-1 checkpoint pathway. N. Engl. J. Med. 375: 1767–1778. https://doi.org/10.1056/NEJMra1514296
- Catalano I, Grassi E, Bertotti A, Trusolin L (2019) Immunogenomics of colorectal tumors: facts and hypotheses on an evolving saga. Trends Cancer 5: 779–788. https://doi.org/10.1016/j.tre-can.2019.10.006
- Chen EX, Jonker DJ, Loree JM, Kennecke HF, Berry SR, Couture F, Ahmad ChE, Goffin JR, Kavan P, Harb M, Colwell B, Samimi S, Samson B, Abbas T, Aucoin N, Aubin F, Koski SL, Wei A Ch-C, Tu D, O'Callaghan ChJ (2019) CCTG CO.26: Updated analysis and impact of plasma-detected microsatellite stability (MSS) and tumor mutation burden (TMB) in a phase II trial of durvalumab (D) plus tremelimumab (T) and best supportive care (BSC) versus BSC alone in patients (pts) with refractory metastatic colorectal car-

- cinoma (rmCRC). J. Clin. Oncol. **37** (suppl: abstr 3512). https://doi.org/10.1200/JCO.2019.37.15_suppl.3512
- Conciatori F, Bazzichetto C, Falcone I, Pilotto S, Bria E, Cognetti F, Milella M, Ciuffreda L (2018) Role of mTOR signaling in tumor microenvironment: an overview. *Int. J. Mol. Sci.* 19. https://doi.org/10.3390/ijms19082453
- Cortez MA, Ivan C, Valdecanas D, Wang X, Peltier HJ, Ye Y, Araujo L, Carbone DP, Shilo K, Giri DK, Kelnar K, Martin D, Komaki R, Gomez DR, Krishnan S, Calin GA, Bader AG, Welsh JW (2015) PDL1 Regulation by p53 via miR-34. J. Natl. Cancer Inst. 108. htt-ps://doi.org/10.1093/jnci/djv303
- Crowell JA, Steele VE, Fay JR (2007) Targeting the Akt protein kinase for cancer chemoprevention. *Mol. Cancer Ther.* **6**: 2139–2148. https://doi.org/10.1158/1535-7163.MCT-07-0120
- Da L, Teng Y, Wang N, Zaguirre K, Liu Y, Qi Y, Song F (2020) Organ-specific immune-related adverse events associated with immune checkpoint inhibitor monotherapy versus combination therapy in cancer: a meta-analysis of randomized controlled trials. Front. Pharmacol. 10: 1671. https://doi.org/10.3389/fphar.2019.01671
- macol. 10: 1671. https://doi.org/10.3389/fphar.2019.01671

 Davis RE, Ngo VN, Lenz G, Tolar P, Young RM, Romesser PB, Kohlhammer H, Lamy L, Zhao H, Yang Y, Xu W, Shaffer AL, Wright G, Xiao W, Powell J, Jiang JK, Thomas CJ, Rosenwald A, Ott G, Muller-Hermelink HK, Gascoyne RD, Connors JM, Johnson NA, Rimsza LM, Campo E, Jaffe ES, Wilson WH, Delabie J, Smeland EB, Fisher RI, Braziel RM, Tubbs RR, Cook JR, Weisenburger DD, Chan WC, Pierce SK, Staudt LM (2010) Chronic active B-cell receptor signaling in diffuse large B-cell lymphoma. Nature 463: 88–92. https://doi.org/10.1038/nature08638
- De la Iglesia N, Konopka G, Lim KL, Nutt CL, Bromberg JF, Frank DA, Mischel PS, Louis DN, Bonni A (2008) Deregulation of a STAT3-interleukin 8 signaling pathway promotes human glioblastoma cell proliferation and invasiveness. *J. Neurosci.* 28: 5870–5878. https://doi.org/10.1523/INEUROSCI.5385-07.2008
- https://doi.org/10.1523/JNEUROSCI.5385-07.2008
 Deppert W, Bruns M (2016) Cancer immunotherapy: weak beats strong. *Aging* (Albany NY) 8: 2607–2608. https://doi.org/10.18632/aging.101134
- Dorand RD, Nthale J, Myers JT, Barkauskas DS, Avril S, Chirieleison SM, Pareek TK, Abbott DW, Stearns DS, Letterio JJ, Huang AY, Petrosiute A (2016) Cdk5 disruption attenuates tumor PD-L1 expression and promotes antitumor immunity. *Science* **353**: 399–403. https://doi.org/10.1126/science.aae0477
- Eng C, Kim TW, Bendell J, Argilés G, Tebbutt NC, Di Bartolomeo M, Falcone A, Fakih M, Kozloff M, Segal NH, Sobrero A, Yan Y, Chang I, Uyei A, Roberts L, Ciardiello F, IMblaze370 Investigators (2019) Atezolizumab with or without cobimetinib versus regorafenib in previously treated metastatic colorectal cancer (IMblaze370): a multicentre, open-label, phase 3, randomised, controlled trial. Lancet Oncol. 20: 849–861. https://doi.org/10.1016/S1470-2045(19)30027-0
- Feld E, Horn L (2016) Targeting PD-L1 for non-small-cell lung cancer. Immunotherapy 8: 747–758. https://doi.org/10.2217/imt-2016-0012
- Green MR, Monti S, Rodig SJ, Juszczynski P, Currie T, O'Donnell E, Chapuy B, Takeyama K, Neuberg D, Golub TR, Kutok JL, Shipp MA (2010) Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. Blood 116: 3268–3277. https://doi.org/10.1182/blood-2010-05-282780
- Grzywnowicz M, Giannopoulos K (2012) The role of receptor programmed death-1 and its ligands in immune system and tumors. Acta Haematol. Polon. 43: 132–145 (in Polish)
- Hamanishi J, Mandai M, Iwasaki M, Okazaki T, Tanaka Y, Yamaguchi K, Higuchi T, Yagi H, Takakura K, Minato N, Honjo T, Fujii S (2007) Programmed cell death 1 ligand 1 and tumor-infiltrating CD8+ T lymphocytes are prognostic factors of human ovarian cancer. *Proc. Natl. Acad. Sci. USA* 104: 3360–3365. https://doi.org/10.1073/pnas.0611533104
 Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, Sosman JA, McDermott DF, Powderly JD, Gettinger SN, Kohrt HE, Horn L, Lawrence DP, Rost S, Leabman M, Xiao Y, Mokatrin A, Canada H, Handa DS, Mellara L, Chan DS, Hall JS (2014)
- Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, Sosman JA, McDermott DF, Powderly JD, Gettinger SN, Kohrt HE, Horn L, Lawrence DP, Rost S, Leabman M, Xiao Y, Mokatrin A, Koeppen H, Hegde PS, Mellman I, Chen DS, Hodi FS (2014) Predictive correlates of response to the anti-PD-L1 antibody MP-DL3280A in cancer patients. Nature 515: 563–567. https://doi.org/10.1038/nature14011
- Holmgaard RB, Zamarin D, Munn DH, Wolchok JD, Allison JP (2013) Indoleamine 2,3-dioxygenase is a critical resistance mechanism in antitumor T cel immunotherapy targeting CTLA-4. J. Exp. Med. 210: 1389–1402. https://doi.org/10.1084/iem.20130066
- Med. 210: 1389–1402. https://doi.org/10.1084/jem.20130066

 Horton BL, Gajewski TF (2018) Back from the dead: TIL apoptosis in cancer immune evasion. Br. J. Cancer 118: 309–311. https://doi.org/10.1038/bjc.2017.483
- Huynh A, DuPage M, Priyadharshini B, Sage PT, Quiros J, Borges CM, Townamchai N, Gerriets VA, Rathmell JC, Sharpe AH, Bluestone JA, Turka LA (2015) Control of PI(3) kinase in Treg cells maintains homeostasis and lineage stability. Nat. Immunol. 16: 188–196. https://doi.org/10.1038/ni.3077

- https://www.nccn.org/patients/guidelines/colon/files/assets/com-
- mon/downloads/files/colon.pdf Ishida Y, Agata Y, Shibahara K, Honjo T (1992) Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. EMBO J. 11: 3887-3895. https://doi. org/10.1002/j.1460-2075.1992.tb05481.x
- Ji RR, Chasalow SD, Wang L, Hamid O, Schmidt H, Cogswell J, Alaparthy S, Berman D, Jure-Kunkel M, Siemers NO, Jackson JR, Shahabi V (2012) An immune-active tumor microenvironment favors clinical response to ipilimumab. Cancer Immunol. Immunother. 61: 1019–1031. https://doi.org/10.1007/s00262-011-1172-6
- Lastwika KJ, Wilson W, Li QK (2016) Control of PD-L1 expression by oncogenic activation of the AKT-mTOR pathway in non-small cell lung cancer. Cancer Res. 76: 227–238. https://doi.org/10.1158/0008-
- Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, Skora AD, Luber BS, Azad NS, Laheru D, Biedrzycki B, Done-hower RC, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Duffy SM, Goldberg RM, de la Chapelle A, Koshiji M, Bhaijee F, Huebner T, Hruban RH, Wood LD, Cuka N, Pardoll DM, Papadopoulos N, Kinzler KW, Zhou S, Cornish TC, Taube JM, Anders RA, Eshleman JR, Vogelstein B, Diaz LA Jr (2015) PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N. Engl. J. Med. 372: 2509–2520. https://doi.org/10.1056/NEJMoa1500596
- Lee J, Kefford R, Carlino M (2016) PD-1 and PD-L1 inhibitors in melanoma treatment: Past success, present application and future challenges. *Immunotherapy* 8: 733–746. https://doi.org/10.2217/imt-
- Lynch D, Murphy A (2016) The emerging role of immunotherapy in colorectal cancer. *Ann. Transl. Med.* **4**: 305. https://doi. org/10.21037/atm.2016.08.29
- Maleki VS, Garrigós C, Duran I (2017) Biomarkers of response to PD-1/PD-L1 inhibition. Crit. Rev. Oncol. Hematol. 116: 116-124. https://doi.org/10.1016/j
- Marzec M, Zhang Q, Goradia A, Raghunath PN, Liu X, Paessler M, Wang HY, Wysocka M, Cheng M, Ruggeri BA, Wasik MA (2008) Oncogenic kinase NPM/ALK induces through STAT3 expression of immunosuppressive protein CD274 (PD-L1, B7-H1). Proc. Natl. Acad. Sci. USA 105: 20852-72085. https://doi.org/10.1073/ pnas.0810958105
- Maxwell PJ, Coulter J, Walker SM, McKechnie M, Neisen J, McCabe N, Kennedy RD, Salto-Tellez M, Albanese C, Waugh DJ (2013) Potentiation of inflammatory CXCL8 signalling sustains cell survival in PTEN-deficient prostate carcinoma. Eur. Urol. 64: 177-188. httos://doi.org/10.1016/j.eururo.2012.08.032
- Nishimura H, Honjo T (2001) PD-1: an inhibitory immunoreceptor involved in peripheral tolerance. Trends Immunol. 22: 265-268. https:// doi.org/10.1038/cmi.2010.28
- O'Donnell JS, Massi D, Teng MWL, Mandala M (2017) PI3K-AKTmTOR inhibition in cancer immunotherapy, redux. Semin. Cancer Biol. 48: 91–103. https://doi.org/10.1016/j.semcancer.2017.04.015
- Oestreich KJ, Yoon H, Ahmed R, Boss JM (2008) NFATc1 regulates PD-1 expression upon T cell activation. J. Immunol. 181: 4832-4839. https://doi.org/0.4049/jimmunol.181.7.4832
- Onishi H, Morisaki T, Katano M (2014) Immunotherapy approaches targeting regulatory T-cells. *Anticancer Res.* **32**: 997–1003. PMID: 22399623
- Parsa AT, Waldron JS, Panner A, Crane CA, Parney IF, Barry JJ, Cachola KE, Murray JC, Tihan T, Jensen MC, Mischel PS, Stokoe D, Pieper RO (2007) Loss of tumor suppressor PTEN function increases B7-H1 expression and immunoresistance in glioma. Nat.
- Med. 13: 84–88. https://doi.org/10.1038/nm1517 Riella LV, Paterson AM, Sharpe AH, Chandraker A (2012) Role of the
- PD-1 pathway in the immune response Am. J. Transplant. 12: 2575–2587. https://doi.org/10.1111/j.1600-6143.2012.04224.x

 Rom-Jurek EM, Kirchhammer N, Ugocsai P (2018) Regulation of programmed death ligand 1 (PD-L1) expression in breast cancer cell lines in vitro and in immunodeficient and humanized tumor mice. Int J. Mol. Sci. 19: https://doi.org/10.3390/ijms19020563
- Smith KM, Desai J (2018) Nivolumab for the treatment of colorectal cancer. Expert. Rev. Anticancer Ther. 18: 611–618. https://doi.org/10.1080/14737140.2018.1480942
- Shin DS, Zaretsky JM, Escuin-Ordinas H, Garcia-Diaz A, Hu-Lieskovan S, Kalbasi Å, Grasso CS, Hugo W, Sandoval S, Torrejon DY, Palaskas N, Rodriguez GA, Parisi G, Azhdam A, Chmielowski B,

- Cherry G, Seja E, Berent-Maoz B, Shintaku IP, Le DT, Pardoll DM, Diaz LA Jr, Tumeh PC, Graeber TG, Lo RS, Comin-Anduix B, Ribas A (2017) Primary resistance to PD-1 blockade mediated by JAK1/2 mutations. Cancer Discov. 7: 188-201. https://doi. org/10.1158/2159-8290.CD-16-1223
- Sobral-Leite M, Salomon I, Opdam M (2019) Cancer-immune interactions in ER-positive breast cancers: PI3K pathway alterations and tumor-infiltrating lymphocytes. Breast Cancer Res. 21: 90. https://doi. org/10.1186/s13058-019-1176-2
- Tapia Rico G, Price TJ (2018) Atezolizumab for the treatment of colorectal cancer: the latest evidence and clinical potential. Expert. Opin. Biol. Ther. 18: 449–457. https://doi.org/10.1080/14712598.20
- Topalian SL, Sznol M, McDermott DF, Kluger HM, Carvajal RD, JD, Leming PD, Lipson EJ, Puzanov I, Smith DC, Taube JM, Wigginton JM, Kollia GD, Gupta A, Pardoll DM, Sosman JA, Hodi FS (2014) Survival, durable tumor remission, and long-term safety in patients with advanced melanoma receiving nivolumab. *J. Clin. Oncol.* **32**: 1020–1030. https://doi.org/10.1200/JCO.2013.53.0105
- Turnis ME, Andrews LP, Vignali DA (2015) Inhibitory receptors as targets for cancer immunotherapy. *Eur. J. Immunol.* **45**: 1892–1905.
- https://doi.org/10.1002/eji.201344413 Valentini AM, Di Pinto F, Cariola F, Guerra V, Giannelli G, Caruso ML, Pirrelli M (2018) PD-L1 expression in colorectal cancer defines three subsets of tumor immune microenvironments. *Oncotarget.* **9**: 8584–8596. https://doi.org/10.18632/oncotarget.24196 Wang T, Niu G, Kortylewski M, Burdelya L, Shain K, Zhang S, Bhat-
- tacharya R, Gabrilovich D, Heller R, Coppola D, Dalton W, Jove R, Pardoll D, Yu H (2004) Regulation of the innate and adaptive immune responses by Stat-3 signaling in tumor cells. *Nat. Med.* **10**: 48–54. https://doi.org/10.1038/nm976
- Wang Q, Liu F, Liu L (2017) Prognostic significance of PD-L1 in solid tumor: An updated meta-analysis. Medicine (Baltimore). 96: e6369. https://doi.org/10.1097/MD.00000000000006369
- Williams L, Bradley L, Smith A, Foxwell B (2004) Signal transducer and activator of transcription 3 is the dominant mediator of the anti-inflammatory effects of IL-10 in human macrophages. J. Immunol. 172: 567–76. https://doi.org/10.4049/jimmunol.172.1.56
- Własiuk P, Putowski M, Giannopoulos K (2016) PD1/PD1L pathway, HLA-G and T regulatory cells as new markers of immunosuppression in cancers (2016) Postepy Hig. Med. Dosw. (online) 0:1044-1058 (in Polish)
- Xiang X, Yu PC, Long D, Liao XL, Zhang S, You XM, Zhong JH, Li LQ (2017) Prognostic value of PD-L1 expression in patients with primary solid tumors. Oncotarget. 9: 5058–5072. https://doi.org/10.18632/oncotarget.23580
- Xue G, Zippelius A, Wicki A, Mandalà M, Tang F, Massi D, Hemmings BA (2015) Integrated Akt/PKB signaling in immunomodulation and its potential role in cancer immunotherapy. J. Natl. Cancer Inst. 107. https://doi.org/10.1093/jnci/djv171
- Yu H, Kortylewski M, Pardoll D (2007) Crosstalk between cancer and immune cells: role of STAT3 in the tumour microenvironment. Nat. Rev. Immunol. 7: 41-51. https://doi.org/10.1038/nri199
- Zeng H (2017) mTOR signaling in immune cells and its implications for cancer immunotherapy. *Cancer Lett.* **408**: 182–189. https://doi.org/10.1016/j.canlet.2017.08.038
- Zhang M, Li G, Wang Y, Wang Y, Zhao S, Haihong P, Zhao H, Wang Y (2017) PD-L1 expression in lung cancer and its correlation with driver mutations: a meta-analysis. Sci Rep. 7: 10255. https:// doi.org/10.1038/s41598-017-10925-
- Zhang P, Bao Z, Xu I, Zhou J, Lu G, Yao Y, Liu R, Gao Q, Shen Y, Zhou J (2017) PD-L1 expression indicates favorable prognosis for advanced lung adenocarcinoma patients treated with pemetrexed. Oncotarget. 8: 66293-66304. https://doi.org/10.18632/oncotarget.19973
- Zhang X, Schwartz JC, Guo X, Bhatia S, Cao E, Lorenz M, Cammer M, Chen L, Zhang ZY, Edidin MA, Nathenson SG, Almo SC (2004) Structural and functional analysis of the costimulatory receptor programmed death-1. Immunity 20: 337-347. https://doi. org/10.1016/s1074-7613(04)00051-2
- Zhuan-Sun Y, Huang F, Feng M, Zhao X, Chen W, Zhu Z, Zhang S (2016) Prognostic value of PD-L1 overexpression for pancreatic cancer: evidence from a meta-analysis. Onco. Targets Ther. 10: 5000-5012. https://doi.org/10.2147/OTT.S146383