

## Anticoagulative effect of pepsin<sup>\*</sup>

Marek Gacko, Wiesława Roszkowska-Jakimiec and Joanna Jasielczuk<sup>✉</sup>

*Departments of Instrumental Analysis and Department of Vascular Surgery, Medical Academy, Białystok, Poland*

Received: 05 July, 1999

**Key words:** pepsin, anticoagulative action

**Anticoagulative effect of pepsin is observed *in vitro* when its concentration is 36  $\mu$ M and higher. This effect is due to inhibition of fibrin monomer polymerization. Protamine abolishes anticoagulative effect of pepsin. Pepsin does not influence platelet aggregation induced by ADP and collagen.**

Anticoagulative effect of heparin and other glycosaminoglycans is connected with their polyanion character [1, 2]. Hence it seemed probable that pepsin with the isoelectric point at pH 1.0 may also demonstrate such an effect [3].

The aim of this report was to show the influence of pepsin on prothrombin, recalcification, celite-cephalin and thrombin times and on the amidolytic activity of thrombin. Also the influence of pepsin on platelet aggregation induced by adenosine 5'-diphosphate and collagen was examined.

### REAGENTS AND METHODS

Adenosine 5'-diphosphate (ADP), fibrinogen, protamine, thrombin, Sigma (St. Louis,

U.S.A.); collagen, Hormonchemie (Munich, Germany); pepsin A, Serva (Heidelberg, Germany); thromboplastin (thromborel S), Behringwerke A.G. (Marburg, Germany); H-D-Phe-Pip-Arg-pNA, Kabi Vitrum (Sweden); cephalite, bioMerieux (Marcy-l'Etoile, France).

Effect of pepsin on prothrombin, recalcification and celite-cephalin times was investigated in the system containing: 0.18 ml of human citrated plasma and 0.02 ml of pepsin (0.09 to 2.88 mM) or 0.02 ml NaCl (0.15 M) in control. To measure prothrombin time 0.2 ml of thromboplastin was added. Recalcification time was measured after addition of 0.2 ml of CaCl<sub>2</sub> (0.025 M). Celite-cephalin time was investigated after addition of 0.1 ml of cephalite and 0.1 ml of CaCl<sub>2</sub> (0.05 M). All measurements of clotting time were made at 37°C.

<sup>\*</sup>This paper was presented as a preliminary communication at the 34th Meeting of the Polish Biochemical Society.

<sup>✉</sup>Corresponding author: Wiesława Roszkowska-Jakimiec, Department of Instrumental Analysis, Medical Academy, P.O.Box 24, 15-230 Białystok 8, Poland; phone: (48) 742 2021, ext. 2361; fax: (48) 8542 4907.

Effect of pepsin on thrombin time was examined in the system containing: 0.18 ml of fibrinogen (0.25%) and 0.02 ml of pepsin (0.09 to 2.88 mM) or 0.02 ml NaCl (0.15 M) in control. After adding 0.2 ml of thrombin (8 U/ml) clotting time was measured at 37°C.

The effect of pepsin on the amidolytic activity of thrombin was investigated in a system [4] containing: 0.65 ml Tris/HCl buffer (0.1 mM, pH 8.4), 0.05 ml of pepsin (0.09 to 2.88 mM) or 0.05 ml NaCl (0.15 M) in control, 0.2 ml of H-D-Phe-Pip-Arg-pNA (0.75 mM) and 0.1

ml of thrombin (5 U/ml). Samples were incubated for 10 min at 37°C. The reaction was stopped by adding 0.1 ml 40% acetic acid and the absorbance was measured at 405 nm. The amount of released pNA was read from a calibrated graph prepared with the use of a standard solutions of this compound.

## RESULTS

Pepsin prolongs prothrombin, recalcification and celite-cephalin times in a concentration-dependent manner (Table 1).

Table 1. Influence of pepsin on prothrombin time, recalcification time and celite-cephalin time

Pepsin $\mu\text{M}$	Prothrombin time (s)	Recalcification time (s)	Celite-cephalin time (s)
0.0	15.3	210.7	75.0
4.5	18.3	210.3	75.3
9.0	22.0	215.0	75.0
18.0	28.3	230.7	75.7
36.0	36.0	248.3	84.3
72.0	42.3	312.7	92.7
144.0	50.0	365.0	162.0

ml of thrombin (5 U/ml). Samples were incubated for 10 min at 37°C. The reaction was stopped by adding 0.1 ml 40% acetic acid and the absorbance was measured at 405 nm. The amount of released pNA was read from a calibrated graph prepared with the use of a standard solutions of this compound.

Neutralization of the anticoagulative effect of pepsin by protamine was investigated in the systems containing 0.18 ml of fibrinogen (0.25%), 0.02 ml of pepsin (0.36 mM), 0.2 ml of thrombin (8 U/ml) and alternatively 0.02 ml of protamine (0.18, 0.36, 0.72 mM) or 0.02 ml NaCl (0.15 M). Clotting time was measured at 37°C.

The influence of pepsin on blood platelet aggregation was investigated in the systems [5] containing: a) 0.9 ml of porcine platelet rich plasma, 0.1 ml of pepsin (0.36 mM) and alternatively 0.1 ml ADP (100 mM) or 0.1 ml of collagen (100  $\mu\text{g}/\text{ml}$ ) and transmittance was mea-

ured; b) 0.9 ml of porcine platelet rich plasma, 0.05 ml of pepsin (0.72 mM) and alternatively 0.05 ml of ADP (200 mM) or 0.05 ml collagen (200  $\mu\text{g}/\text{ml}$ ) and transmittance was measured.

Also pepsin prolongs thrombin clotting time (Table 2). However, it does not inhibit the amidolytic activity of thrombin against H-D-Phe-Pip-Arg-pNA.

Protamine at 18  $\mu\text{M}$  concentration decreases and at 36  $\mu\text{M}$  completely abolishes the prolongation of thrombin time by 0.36 mM pepsin (Table 3).

Table 2. Influence of pepsin on thrombin time and amidolytic activity of thrombin

Pepsin $\mu\text{M}$	Thrombin time (s)	pNA $\mu\text{M}$
0.0	16.3	108.3
4.5	16.7	108.3
9.0	16.7	108.3
18.0	16.7	107.7
36.0	26.3	108.3
72.0	34.0	109.0
144.0	41.3	108.7

Table 3. Neutralization of anticoagulative effect of pepsin by protamine

Test components	Protamine $\mu\text{M}$			
	0.0	9.0	18.0	36.0
	Clotting time (s)			
Protamin + NaCl	15.0	15.3	15.7	15.7
Pepsin* + protamine	26.7	26.3	16.7	15.3

\* Pepsin concentration was  $36 \mu\text{M}$

Pepsin does not induce blood platelet aggregation and does not inhibit platelet aggregation induced by ADP and collagen.

## DISCUSSION

The presented data show that pepsin prolongs prothrombin, recalcification, celite-celphalin and thrombin times. In these tests the generated or added thrombin induces the formation of fibrin monomers, their polymerization and formation of fibrin clot. The lack of inhibition of the amidolytic activity of thrombin excludes the effect of pepsin. It allows to suppose that the anticoagulative effect of pepsin depends on the formation of complexes with fibrin monomers thus making their polymerization difficult. The abolishment of the anticoagulative effect of pepsin by protamine with which it forms stable complexes supports this supposition [6]. It is also confirmed by the inhibition of polymerization of isolated fibrin monomers by pepsin (unpublished). Polymerization of fibrin monomers is also inhibited by fibrinogen degradation products [7], haemoglobin [8] and some synthetic polyamino acids [9].

An impairment of haemostasis manifested by persistent bleedings and haemorrhages difficult to control is observed in some pathologic conditions affecting gastric circulation [10, 11]. This pathology may result from the anticoagulative effect of hydrochloric acid [11, 12], the increased fibrinolytic activity of

gastric blood vessels [13, 14], and perhaps the anticoagulative effect of pepsin as well. About 1% of pepsin synthesized in the principal cells of gastric mucosa gets into circulating blood where its concentration in general circulation is from 1.5 to 5.0 nM [15]. Local concentration of pepsin in gastric circulation is probably much higher. Blood flowing from blood vessels into stomach lumen will get into contact with pepsin of the gastric juice. Concentration of pepsin in the gastric juice may surpass  $100 \mu\text{M}$  [16].

## REFERENCES

1. Bourin, M.C. & Lindahl, U. (1993) Glycosaminoglycans and the regulation of blood coagulation. *Biochem. J.* **289**, 313-330.
2. Undas, A. & Gajewski, P. (1998) Role of heparinoids in treatment of vascular diseases. *Pol. Arch. Med. Wewn.* **99**, 430-435 (in Polish).
3. Perlman, G.E. (1970) Porcine pepsinogen and pepsin; in *Structure - Function Relationship of Proteolytic Enzymes* (Desnuelle, P., Neurath, H. & Ottesen, M., eds.) pp. 261-270, Munksgaard, Copenhagen.
4. Lottenberg, R., Hall, J.A., Fenton, J.W. & Jackson, C.M. (1982) The action of thrombin on peptide *p*-nitroanilide substrates: Hydrolysis of Tos-Gly-Pro-Arg-pNA and D-Phe-Pip-Arg-pNA by human  $\alpha$ - and  $\gamma$ - and bovine  $\alpha$ - and  $\beta$ -trypsin. *Thrombos. Res.* **28**, 313-322.

5. Born, G.V.R. (1963) Quantitative investigation into aggregation of blood platelets. *J. Physiol.* **168**, 178-195.
6. Roszkowska-Jakimiec, W., Fouad, Khamis A., Gacko, M. & Worowska, A. (1996) Properties of pepsin as a polyanion protein. Part I: Complexes of pepsin with protamine. *Bromat. Chem. Toksykol.* **29**, 381-386 (in Polish).
7. Lipiński, B., Węgrzynowicz, Z., Budzyński, A.Z., Kopeć, M., Latało, Z.S. & Kowalski, E. (1967) Soluble unclotable complexes formed in the presence of fibrinogen degradation products (FDP) during the fibrinogen-fibrin conversion and their potential significance in pathology. *Thrombos. Diathes. Haemorrhag.* **17**, 65-69.
8. Brezniak, D.V., Moon, D.G., Beaver, J.A. & Fenton, J.W. (1994) Haemoglobin inhibition of fibrin polymerization and clotting. *Blood Coagulat. Fibrinol.* **5**, 139-143.
9. Carr, M.E., Cromartie, R. & Gabriel, D.A. (1989) Effect of homopoly(L-amino acids) on fibrin assembly: Role of charge and molecular weight. *Biochemistry* **28**, 1384-1388.
10. Vellacott, K.D., Drenfield, M.W., Atkinson, M. & Langman, M.J.S. (1982) Comparison of surgical and medical management of bleeding peptic ulcer. *Br. Med. J.* **284**, 548-550.
11. Sajewicz, Z., Poniewierka, E., Lubczyńska-Kowalska, W., Szmigielski, R. & Bitter, J. (1996) Influence of gastric juice and sucralfate on fibrinolysis. *Ter. Lek.* **24**, 142-144 (in Polish).
12. Green, F.W., Kaplan, M.M., Curtis, L.E. & Levine, I.H. (1978) Effect of acid and pepsin on blood coagulation and platelet aggregation. A possible contribution to prolonged gastroduodenal mucosal hemorrhage. *Gastroenterology* **74**, 38-43.
13. Prokopowicz, J., Gabryelewicz, A., Wołosowicz, N., Szalaj, W. & Puchalski, Z. (1975) Peripheral and gastric fibrinolysis in patients with gastric carcinoma. *Z. Exp. Chirurg.* **8**, 309-312.
14. O'Brien, T.E., Handley, H. & Irving, M.H. (1979) Fibrinolytic activity in gastric venous blood. A comparison between normal and diseased stomach. *Gastroenterology* **76**, 509-514.
15. Samloff, I.M. (1982) Pepsinogen I and II: Purification from gastric mucosa and radioimmunoassay in serum. *Gastroenterology* **82**, 26-33.
16. Taylor, W.H. (1986) Advances in the biochemistry and pathophysiology of pepsin 1. *Z. Med. Lab. Diagn.* **27**, 115-120.