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Osteosarcoma's genetic landscape painted by genes' mutations

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Purpose: Osteosarcoma (OS) is one of the most common primary bone tumors. Direct pathogenesis remains unknown, however, genes' mutations are proven to participate in the process. This study aimed to examine the most frequently mutated genes in OS to appoint candidates for the cancer markers. Methods: Using the COS-MIC Catalogue twenty the most frequently mutated genes were selected leading to an up-to-date genetic OS landscape summary. The genes can be classified into four categories: suppressor genes (TP53, RB1, NCOR1, SMAD2, NF1, TSC2, KMT2C), proto-oncogenes (GNAS, BRAF, MLLT3), epigenetic and post-translational modification-related genes (SMARCA4, ARID1A, ATRX, BCOR, H3F3A) and cell growth and survival regulating genes (EGFR, CAMTA1, LRP1B, PDE4DIP, MED12). Results and conclusions: Their role in cancerogenesis was confirmed by the analysis of available articles published previously. The results of the study indicate that examination of selected genes' mutations might help to identify patients' predisposition to OS development, as well as monitor the disease progression, and establish prognosis. However, to fully understand the pathogenesis of OS further studies are required.

Keywords: osteosarcoma, genes'mutations, COSMIC catalogue, biomarkers

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Abbreviations: ALP, alkaline phosphatase; ANPEP, alanyl aminopeptidase; AP-1, activator protein 1; ARID1A, AT-rich interactive domaincontaining protein 1A; ATRX, alpha thalassemia/mental retardation syndrome X-linked; BCOR, BCL6 Corepressor; BRAF, serine/threonine kinase; c-Myc, cellular myelocytomatosis oncogene; CAMTA1, calmodulin binding transcription activator 1; CD133, CD133 antigen; COSMIC, Catalogue Of Somatic Mutations In Cancer; CRP, C-reactive protein; EGFR, epidermal growth factor receptor; EMT, epithelial-mesenchymal transition; FLNA, filamin A; FUCA1, alpha-L-fucosidase 1; GNAS, guanine nucleotide binding protein alpha stimulating; H3F3A, H3 histone family 3A; HER-4, human epidermal growth factor receptor 4; ICB, immune checkpoint blockade; ICIs, immune checkpoint inhibitors; KTM2C, lysine methyltransferase 2C; LAMA3, laminin subunit alpha 3; LGALS1, galectin 1; LRP1B gene, low density lipoprotein receptor-related protein 1B; MATN3, matrilin 3; MED12, mediator complex subunit 12; miRNA, microRNA; MLLT3, MLLT3 super elongation complex subunit 3; MMGL, myomegalinlike; MSCs, mesenchymal stem cells; NANOG, homeobox protein NA-NOG; NCOR1, nuclear receptor corepressor 1; NF1, neurofibromin 1; OS, Osteosarcoma; PDE4DIP, phospodiestrase 4D-interacting protein; Rb, retinoblastoma protein; RB1, RB transcriptional corepressor 1; SEC, super elongation complex; SGCG, sarcoglycan gamma; SMAD2, SMAD family member 2; SMARCA4, SWI/SNF-related, matrix-associated, actin-dependant regulator of chromatin, subfamily a, member 4; SOX4, SRY-box transcription factor 4; TCGA, The Cancer Genome Atlas; TICs, tumor-initiating cells; TP53, tumor protein p53; TSC, tu-berous sclerosis complex; TSC2, TSC complex subunit 2; TWIST, Twist Family BHLH Transcription Factor; VEGFA,Vascular endothelial growth factor A; WNT5A, Wnt Family Member 5A

INTRODUCTION

Osteosarcoma (OS) is one of the most common primary bone tumors. It occurs intraosseous, mainly in the metaphyseal region of the long bones. OS is inflicting an accelerated osteoid matrix production, which is connected to being developed at sites where bone grows expeditiously. Current data prove that OS has two main peaks of incidence: in childhood and adolescence. OS is the third most common type of tumor affecting young people, with the highest incidents in the early twenties (Wu & Livingston, 2020; Sun *et al.*, 2020; Misaghi *et al.*, 2018; Czarnecka *et al.*, 2020).

Osteosarcoma is divided into several subtypes (Fig. 1). It can be categorized depending on the region of bone it affects (the surface and central part of the bone, and within the medulla). The staging scheme divides OS into two classes depending on the grade of the tumor (low or high), as well as the localization of the compartment (extra- or intra-compartmental). Additionally, it is classified considering the metastatic level of the tumor. The most common classification groups OS as osteoblastic, chondroblastic, fibroblastic, and small cell subtypes. Among mention above types of ostesarcoma, the most often diagnosed one is a high-grade tumor, occurring extra-cortically (Misaghi *et al.*, 2018).

Even though the exact mechanisms responsible for OS pathogenesis are not known, it is confirmed that genetics play a role in the tumor origin. The most common characteristic of OS includes genome disorganization, together with alterations of tumor suppressor and DNA repair, as well as changes in cell cycle aneuploids with chromosomal alterations (de Azevedo *et al.*, 2020).

Classification of osteosarcoma subtypes

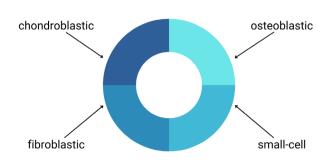


Figure 1. The most common classification of osteosarcoma subtypes, depending on the predominant type of cells affected by OS. It was proved that OS can be developed as a consequence of alteration in tumor suppressor genes, including *TP53* and *Rb1*, during hereditary disorders such as Li-Fraumeni cancer family syndrome and retinoblastoma (Porter *et al.*, 1992; Fletcher & Unni, 2002; Misaghi *et al.*, 2018; Sun *et al.*, 2020). However, also other genes, such as transcription factors, and tumor suppressor genes, including *CAMTA1*, or *KMT2C* might participate in osteosarcoma development (He *et al.*, 2021; Chen *et al.*, 2016; Chiappetta *et al.*, 2019).

Currently, there are no laboratory tests or specific biomarkers to diagnose osteosarcoma (Misaghi *et al.*, 2018). Even though there are some molecules expressed in OS that are proposed as their potential markers. The most promising one appears to be alkaline phosphatase (ALP), which increased serum level and seems to correlate positively with tumor volume (Limmahakhun *et al.*, 2011). Additionally, C-reactive protein (CRP), Cathepsin D, osteocalcin, SATB2, and aspartic endoprotease can help distinguish OS (Misaghi *et al.*, 2018; Czarnecka *et al.*, 2020; Agustina *et al.*, 2018; Machado *et al.*, 2016; Tallegas *et al.*, 2022).

In addition to proteins, microRNA expression also seems to be valuable in the diagnosis of osteosarcomas. Overexpression of miR-421 and miR-191 proves to be linked with proliferation, migration, and malignant character of the tumors. Their levels in the serum of OS patients were higher compared to samples from healthy volunteers (Czarnecka *et al.*, 2020).

Moreover, the presence of tumor-initiating cells (TICs) may help in OS diagnostics. TICs expressing markers of stem cell phenotype, such as NANOG or SOX4 were observed in both primary and metastatic tumor tissue (Yan *et al.*, 2016). These proteins, together with CD133+, which is a distinctive marker of TICs observed in osteo-sarcoma cell lines (Czarnecka *et al.*, 2020). It has been proved that the expression of CD133 in OS patients is related to distant metastasis and poor prognosis. making this a tumor marker (Xie *et al.*, 2018).

Finally, the prevalence of mesenchymal stem cells (MSCs) connected with OS progression is proposed to be the tumor marker. Changes in the expression of c-Myc, Rb, AP-1 or TWIST were shown to contribute to the transformation of MSCs into osteosarcoma tumor cells (Yang et al., 2020). Most sarcomas characterize a permanent mesenchymal state. However, their phenotype can change by factors regulating epithelial-mesenchymal transition - EMT. These factors have been shown to be highly correlated with the invasiveness and higher risk of metastasis in malignant OS (Wu et al., 2019; Yu, Yustein & Xu, 2021). An EMT-related genes' panel (including LAMA3, LGALS1, SGCG, VEGFA, WNT5A, MATN3, ANPEP, FUCA1, and FLNA) was used as a predictive marker in a multi-cohort study of OS. The panel proved to be a reliable tool for estimating the overall survival of OS patients. Moreover, it was proposed as a selection method for patients with metastases for personalized treatment (Yiqi et al., 2020).

Even though numerous studies describe molecules, whose expression may characterize osteosarcomas, all pointed proteins and/or miRNA are not specific to OS. Thus, further studies are needed to establish biomarkers helping to diagnose and treat OS.

The present study aims to provide a comprehensive, up-to-date study of genes the most frequently mutated in osteosarcoma using the COSMIC Catalogue. Such genes would become candidates for OS markers.

COSMIC, the Catalogue Of Somatic Mutations In Cancer (https://cancer.sanger.ac.uk) is a database providing tools to explore somatic mutations and their correlation with human cancers. The catalogue is the collection of somatic mutations. The data used to build this catalogue derives strictly from the scientific literature. COSMIC additionally provides information about patients' genetic predispositions and environmental factors participating in the process of cancerogenesis (Tate *et al.*, 2019).

MATERIALS AND METHODS

To identify the most common somatic mutations occurring in osteosarcoma patients, the COSMIC Cancer Browser tool (https://cancer.sanger.ac.uk/cosmic/browse/tissue) was used. The catalogue provides information about 282 osteosarcoma samples (classified into 9 histological subtypes). The biological relevance of the 20 most frequently mutated genes pointed out by the COSMIC Cancer Browser was investigated by the analysis of publications available on PubMed (https://pubmed.ncbi.nlm.nih.gov/). To search for the publications, specific keywords, such as: selected genes' name, osteosarcoma, cancer, and mutation were used. For each gene research was divided into two keyword panels – first: selected genes' name, cancer, mutation; second: selected genes' name, osteosarcoma, mutation.

RESULTS AND DISCUSSION

Genes the most frequently mutated in OS

The analysis of the COSMIC Catalogue provided a list of genes in which somatic mutation may participate in osteosarcoma development. Out of 18,309 entries of genes, the top 20 genes, based on the frequency of their

Table 1. The frequency of gene mutation in OS samples in the COSMIC Catalogue

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Gene	Number of OS samples available in COSMIC catalogue	Number of mutated sam- ples and % of frequency
TP53	371	91 (25%)
RB1	285	22 (8%)
ATRX	212	16 (8%)
KMT2C	215	13 (6%)
LRP1B	136	8 (6%)
CAMTA1	134	7 (5%)
NCOR1	208	9 (4%)
MLLT3	134	6 (4%)
PDE4DIP	134	6 (4%)
GNAS	352	9 (3%)
SMARCA4	211	6 (3%)
ARID1A	210	6 (3%)
H3F3A	453	11 (2%)
EGFR	280	5 (2%)
BRAF	257	6 (2%)
SMAD2	243	5 (2%)
NF1	216	5 (2%)
MED12	210	5 (2%)
TSC2	210	5 (2%)
BCOR	209	5 (2%)

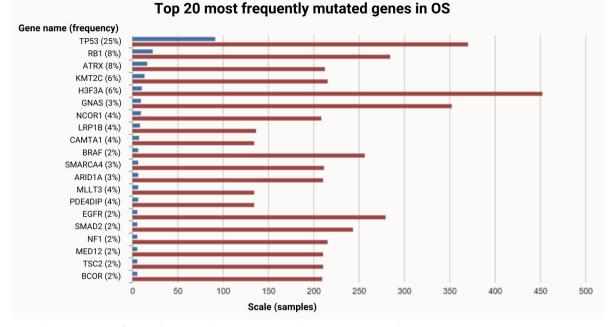


Figure 2. The top 20 most frequently mutated genes in OS according to COSMIC Catalogue

mutation were selected. They are shown in Table 1 and shown in Fig. 2.

All these genes are directly or indirectly involved in products an oncogenesis (Table 2). Seven out of these 20 genes – *TP53, RB1, NCOR1, SMAD2, NF1, TSC2, KMT2C* are protooncog Table 2. Genes the most frequently mutated in osteosarcomas and their function

known to be tumor suppressor genes. Thus, their mutations lead to the expression of malfunctioning protein products and strictly correlate with tumourigenesis. The following three genes: *GNAS*, *BRAF*, and *MLLT3* are protooncogenes. Other genes – *SMARCA4*, *ARID1A*, heir function

Gene	Function	References
TP53	tumor suppression	(Synoradzki <i>et al.,</i> 2021)
RB1	tumor suppression	(Li et al., 2022)
ATRX	epigenetics and post-translational modifi- cations	(He <i>et al.</i> , 2015)
KMT2C	tumor suppression	(Gala et al., 2018; Lian et al., 2022; Liu et al., 2021)
LRP1B	cell growth, metabolism and survival	(Wang & Xiong, 2021)
CAMTA1	cell growth, metabolism and survival	(Lu <i>et al.</i> , 2018)
NCOR1	tumor suppression	(Tang <i>et al.</i> , 2020)
MLLT3	protooncogene	(Sun <i>et al.,</i> 2017)
PDE4DIP	cell growth, metabolism and survival	(Soejima <i>et al.</i> , 2001; Lehnart <i>et al.</i> , 2005)
GNAS	protooncogene	(Nomura <i>et al.</i> , 2014; Zauber, Marotta & Sabbath-Solitare, 2016; Patra <i>et al.</i> , 2018; Afolabi <i>et al.</i> , 2022)
SMARCA4	epigenetics and post-translational modifi- cations	(Xu <i>et al.,</i> 2021; Jelinic <i>et al.,</i> 2014)
ARID1A	epigenetics and post-translational modifi- cations	(Xu <i>et al.,</i> 2019)
H3F3A	post-translational modifications of histone H3.3	(Sturm <i>et al.</i> , 2012; Park <i>et al.</i> , 2016)
EGFR	cell growth, metabolism and survival	(Wang <i>et al.,</i> 2004)
BRAF	protooncogene	(Śmiech <i>et al.,</i> 2020)
SMAD2	tumor suppression	(Piek, Heldin & Ten Dijke, 1999; Pasche, 2001)
NF1	tumor suppression	(Trovó-Marqui & Tajara, 2006)
MED12	cell growth, metabolism and survival	(Ding <i>et al.</i> , 2008)
TSC2	tumor suppression	(Inoki <i>et al.</i> , 2002)
BCOR	epigenetics and post-translational modifi- cations	(Huynh <i>et al.</i> , 2000)

ATRX, BCOR, and H3F3A are related to the epigenetic status of gene expression or posttranslational histone modifications. Finally, EGFR, CAMTA1, LRP1B, PDE4DIP, and MED12 are associated with cell growth, metabolism, and survival.

Molecular basis of OS

Genome destabilization, aneuploids with chromosomal changes, dysregulation of cell cycle and tumor suppressor genes, as well as a lack of DNA repair are some of the most prevalent characteristics of OS (de Azevedo *et al.*, 2020). Epigenetic changes are also recognized as risk factors for OS (Sharma, Kelly & Jones, 2009). Still, the pathogenesis of OS is complex and poorly understood. Thus, the identification of additional genes involved in oncogenesis may lead to advances in understanding OS biology, as well as shortening the time needed to establish diagnosis and introduce proper treatment.

The results of the COSMIC Catalogue analysis allowed distinguishing 20 genes the most frequently mutated in cancer. All recognized genes (Table 2) are known to be key regulators of vital cellular processes such as DNA repair, cell proliferation and differentiation. Thus, their mutations might lead to cancer development.

The most frequently mutated genes and their possible role in OS biology

Among the genes examined during the analysis of the COSMIC Catalogue, the most frequently mutated gene in OS was TP53. Its mutation characterized 25% of samples available in the database. TP53 encodes the P53 protein, which is an essential transcription factor with a tumor suppressor function. It stimulates several processes that result in cell cycle arrest, DNA repair, alterations in metabolism in response to cellular stress, apoptosis, and cell senescence. Moreover, it plays an important role in ontogenesis, myogenesis, and angiogenesis (Synoradzki et al., 2021). Thus, mutated TP53 appears to be essential for cancer development and its progression. Mutations in TP53 are known to increase protein instability, which affects clinical characteristics of osteosarcoma, such as metastatic potential, tumor type, and grade as well as its aggressiveness. According to Chen et al. TP53 mutations are a reliable predictive indicator for osteosarcoma patient survival (Chen et al., 2016).

Another gene commonly mutated in OS is Retinoblastoma transcriptional corepressor 1 gene - RB1. Its mutations were observed in 8% of samples available in the COSMIC catalogue. Between 50% and 70% of osteosarcoma cases exhibit mutations in RB1 (Wu & Livingston, 2020). RB1 encodes retinoblastoma protein controlling the transition from the G1 to S phase. This makes it a crucial regulator of cell cycle progression. It is suggested that during OS oncogenesis, alterations of RB1 may correlate with TP53 inactivation (Wu & Livingston, 2020; Li et al., 2022). Indeed, using mice models it was proved that deletion of RB1 leads to osteosarcomas development. Osteoblast development and mineralization as well as an increase in the expression of osteogenic markers were positively impacted by the gene deletion (Li et al., 2022).

Yet another gene mutated in 8% of OS is *ATRX* – alpha thalassemia/mental retardation syndrome X-linked. The gene encodes the protein that plays a role in chromatin remodeling and DNA repair. Loss of *ATRX* function has been linked to the accumulation of repetitive DNA sequences and alterations in the epigenetic regulation of the gene expression, which may contribute to the

development and progression of tumor (He *et al.*, 2015). Mutations in *ATRX* have been observed in a variety of cancer types (Haase *et al.*, 2018). In osteosarcoma, *ATRX* dysfunction was linked with chromosomal instability, which contributes the tumor development. The gene's mutations were also associated with poor prognosis and resistance to chemotherapy (Masliah-Planchon *et al.*, 2018). Thus, *ATRX* is pointed out as a promising biomarker of osteosarcoma.

6% of OS samples in the COSMIC catalogue manifested mutation of the KTM2C gene. KTM2C has been shown to act as a tumor suppressor. Its expression in OS has been shown to be downregulated, like in a variety of other human cancers, including breast and prostate cancer (Gala et al., 2018; Lian et al., 2022; Liu et al., 2021). Loss of KTM2C expression is associated with poor prognosis and resistance to chemotherapy (Lian et al., 2022; Liu et al., 2021). What is more, KTM2C has been shown to regulate the other tumor suppressor genes, such as TP53 and p16INK4a. By removing methyl groups from histones, KTM2C also activates the expression of oncogenes, such as MYC (Liu et al., 2021; Lian et al., 2022; Gala et al., 2018). All these data demonstrate that the KTM2C gene can play an important role in OS development.

6% of available OS samples of the COSMIC catalogue were mutated in the LRP1B gene - Low Density Lipoprotein Receptor-Related Protein 1B that encodes cell surface receptor for LDL. LRP1B has been previously shown to play a role in tumor growth and progression. Additionally, it regulates angiogenesis in some of the cancer types, such as non-small cell lung cancer, renal cell cancer, and neuroglioma (Wang & Xiong, 2021). The gene overexpression observed in various cancers is linked to poor prognosis and decreased patient survival (Brown et al., 2021; Príncipe et al., 2021; Wang & Xiong, 2021). LRP1B is associated with the occurrence of osteosarcoma and its overexpression correlates with poor prognosis and decreased patient survival. Therefore the gene might be one relevant factor helping in OS diagnosis (Xu et al., 2021).

A significant role in tumorigenesis and tumor progression is also played by the CAMTA1 gene encoding a transcription factor - Calmodulin Binding Transcription Activator 1 (He et al., 2021). The gene down-expression correlates with chemoresistance (Pan et al., 2022). It has been shown that the expression of long noncoding RNA CAMTA1 (IncCAMTA1) in breast cancer affects cells viability and promotes their migratory state. Knock-down of IncCAMTA1 leads to cancer cell apoptosis (Lu et al., 2018). CAMTA1 mutations were observed in 5% of OS COSMIC samples. Even though there is a lack of information about the gene significance for osteosarcoma, still in Epithelioid hemangioendothelioma, that can arise from soft tissues and bones, CAMTA1 was proposed to be an immunohistochemical marker (Anderson & Jo, 2021).

Yet another gene shown to play a role in tumor development and its progression is *NCOR1* (Nuclear Receptor Corepressor 1) gene. COSMIC catalogue analysis showed that as many as 4% of OS samples showed *NCOR1* mutation. The gene encodes a corepressor involved in the regulation of gene expression (Tang *et al.*, 2020). *NCOR1* was observed to be frequently upregulated in breast cancer, and its high expression is associated with good prognosis and better patient survival (Noblejas-López *et al.*, 2018). Interestingly, in bladder cancer, *NCOR1* strongly correlated with immunogenicity, and its mutations were shown to cause activation of anti-tumor immunity, as well as overexpression of immune-related genes (Lin *et al.*, 2021). According to Luo *et al.* NCOR is widely amplificated in osteosarcoma, as it might participate in tumor growth as a regulator of other gene transcription (Luo *et al.*, 2019).

Four percentage of OS samples of the COSMIC catalogue showed mutations of the *MLLT3* gene. MLLT3 protein is part of a super elongation complex (SEC), which is needed to increase the catalytic function of RNA polymerase II (Calvanese *et al.*, 2019). The knockout of the *MLLT3* gene can lead to decreased proliferation of osteosarcoma cell lines, associated with the JNK signaling pathway. Thus, *MLLT3* can be an oncogene candidate involved in osteosarcomas development (Sun *et al.*, 2017).

As much as 4% of samples deposited in the COS-MIC catalogue were characterized by *PDE4DIP* gene mutations. Even though there is no information about the gene significance in osteosarcomas, still, PDE4DIP protein, also known as MMGL (myomegalin-like) protein is involved in several intracellular metabolic pathways, which may control OS biology (Soejima *et al.*, 2001; Lehnart *et al.*, 2005).

Three percentage of OS samples carried a mutation of GNAS (Guanine Nucleotide Binding Protein, Alpha Stimulating). The gene encodes a signaling molecule involved in many cellular processes involving cAMP signaling. It has been shown to play a role in tumorigenesis (Nomura *et al.*, 2014; Zauber, Marotta & Sabbath-Solitare, 2016; Patra *et al.*, 2018; Afolabi *et al.*, 2022). GNASmutations were observed in various cancers, such as colon, pancreas, and gastrin adenocarcinoma as well as in fibrous dysplasia, which is known to have the potential to form malignancies such as osteosarcoma (Sugiura *et al.*, 2018).

Three percentage of samples deposited in the COS-MIC catalogue manifested mutations of the *SMARCA4* gene. *SMARCA4* encodes the catalytic subunit of SWI/ SNF complexes, which is associated with chromatin remodeling and making genes accessible for transcriptional factors. Unfortunately, there is a lack of information about the significance of gene alterations in osteosarcomas.

Also, the *ARID1A* gene product (AT-rich interactive domain-containing protein 1A) is a part of the SWI/ SFI complex, associated with chromatin remodeling (Xu *et al.*, 2019). *ARID1A* mutations were observed in many cancers before (Jones *et al.*, 2010). Recent studies show that in osteosarcomas the expression of *ARID1A* is down-regulated compared to non-tumor tissues (Xu *et al.*, 2019). This can be correlated with the gene mutations measured in 3% of OS samples of the COSMIC catalogue.

Yet another mutated gene affecting the structure of chromatin and contributing to tumorigenesis is H3F3A (Sturm *et al.*, 2012; Park *et al.*, 2016). H3F3A overexpression has been shown to correlate with aggressive phenotype and chemoresistance leading to lower survival rates and relapse in lung cancer patients (Park *et al.*, 2016). COSMIC catalogue analysis showed that 2% of OS samples are mutated in H3F3A. This data finds approval in Koelsche *et al.* study showing hotspot mutations in H3F3A in six osteosarcoma cases analysed by the authors. Observed alternations were correlated with age and increased the incidence of osteosarcoma in patients over the age of 30 (Koelsche *et al.*, 2017).

Two percentage of the samples of the COSMIC catalogue were carrying mutations in the *BCOR* gene. BCOR protein is involved in the process of deacetylation of histones and when over-expressed inhibits BCL-6 tumor suppressor (Huynh *et al.*, 2000). Mutations in *BCOR* are associated with many types of cancer, such as clear cell sarcoma of the kidney, endometrial stromal sarcoma, or small round blue cell sarcomas (Astolfi *et al.*, 2019; Ueno-Yokohata *et al.*, 2015; Marinõ-Enriquez *et al.*, 2018; Sbaraglia *et al.*, 2020).

Among samples deposited in the COSMIC catalogue, approximately 2% carry mutations in the MED12 gene. MED12 is part of a larger, multiprotein complex that mediates polymerase II RNA during the process of transcription (Ding et al., 2008). MED12 is associated with 70% of uterine leiomyomas and fibroepithelial tumors of the breast (Croce & Chibon, 2015; Lerwill et al., 2022). As for now, only two clinical case reports described MED12 mutations in osteosarcomas. Le and others (Le et al., 2021) showed that the patient with MED12-mutated high-grade uterine sarcoma developed osteosarcoma with the same L36R missense mutation. Another team described MED12 mutation in osteosarcoma which further led to the development of breast malignant phyllodes tumor (Tokoyoda et al., 2018). More study is needed to assess the link between MED12 mutations and the occurrence of osteosarcoma, especially since according to The Cancer Genome Atlas (TCGA) data, mutations in MED12 can be biomarkers helping to select patients for the therapy with immune checkpoint inhibitors (ICIs) (Zhou et al., 2022).

Two percentage of OS samples from the COSMIC catalogue displayed changes in the EGFR gene encoding receptor for the epidermal growth factor (EGFR). The receptor is known for its crucial role in the signaling pathway responsible for cell proliferation, differentiation, and/or survival (Wang et al., 2004). Mutations of EGFR in a variety of malignancies are well documented. Most frequent is the gene's amplification leading to EGFR overexpression increasing cellular signaling (Sigismund, Avanzato & Lanzetti, 2018; Cheng et al., 2021). The gene importance for OS has been already documented. Sheng-Lin and others (Sheng-Lin et. al., ????) reported that high expression of EGFR together with HER-4 (human epidermal growth factor receptor 4) was associated with distant metastasis and the level of EGFR expression was proposed to be a potential prognostic biomarker of OS (Wang et al., 2018).

Among OS samples deposited in the COSMIC catalogue, 2% manifested mutations of the SMAD2 gene. This low number of samples can indicate that the gene mutations are not osteosarcoma specific. However, is worth mentioning that SMAD2, as a part of the TGF-beta superfamily, plays an important role in cell signaling, proliferation, and differentiation (Piek et al., 1999) and may act as an oncosuppressor (Piek et al., 1999; Pasche, 2001). Moreover, it has been shown that there are statistically significant changes in SMAD2 expression in OS compared to healthy tissue. The correlation between alterations of Smad signaling, the high proliferation rate, and the invasive phenotype of osteosarcoma cells was observed in OS with Serine/threonine kinase 39 (STK39) knockdown (Won et al., 2010; Yang et al., 2013; Huang et al., 2017).

Another gene selected during our study was the NF1 gene. According to the data from the COSMIC catalogue 2% of OS samples were carrying the gene's mutations. The product of NF1 is neurofibromin 1 – a cytoplasmic protein expressed in neurons, Schwann cells, and leukocytes. The protein is a part of the RAS/MAPK signaling pathway controlling cell proliferation (Huang *et al.*, 2017). It is suggested that NF1 can act also as an oncos-

uppressor. Although the NF1 gene is associated mostly with neural malignancies, its mutations are observed in many cancers, e.g. lung cancers or melanomas (Trovó-Marqui & Tajara, 2006). Further studies are needed to prove NF1 role in OS development.

An additional gene involved in the inhibition of cell proliferation is the tumor-suppressor gene TSC2 (Inoki et al., 2002). The gene's mutations were described in tuberous sclerosis complex (TSC) and non-small cell lung cancer. It was proved that patients with TSC1/TSC2 loss can benefit from target therapy including treatment with immune checkpoint blockade (ICB) (Huang et al., 2022). Kuroda et al. reported a clinical case of a young patient with tuberous sclerosis complex (TSC) who developed osteosarcoma. Analysis of the tumor sample showed additional mutations in TP53 as well as TSC2 genes (Kuroda et al., 2021). This observation is in agreement with COSMIC catalogue data showing that approximately 2% of OS samples were characterized by mutated TSC2.

Two percentage of OS samples were also characterized by mutated proto-oncogene BRAF. BRAF activates signaling pathways involved in the cell cycle, including proliferation, differentiation, and cell death. The gene mutations may result in malignancies development. In fact, dysregulation of gene expression was observed in several cancers, including melanoma, colorectal cancer, and non-small cell lung cancer (Smiech et al., 2020). Pignochino et al. reported that in 4 of 30 osteosarcoma patients analysed in their study, BRAF gene mutation was confirmed (Pignochino et al., 2009).

CONCLUSIONS

COSMIC catalogue analysis allows to identify genes that are the most frequently mutated in osteosarcoma. Since 20 of the selected genes have previously been shown to be involved in cancerogenesis, thus their mutation might also play a role in OS development. However, to prove their clinical significance and the possibility of using them as OS biomarkers further studies are needed. The results of such a study would help to identify patient predisposition to OS development, follow cancer progression, and establish patients' treatment and prognosis.

REFERENCES

- Afolabi HA, Salleh SM, Zakaria Z, Ch'ng ES, Mohd Nafi SN, Ab-dul Aziz AA Bin, Irekeola AA, Wada Y, Al-Mhanna SB (2022) A GNAS gene mutation's independent expression in the growth of colorectal cancer: a systematic review and meta-analysis. Cancers (Basel) 14: 5480. https://doi.org/10.3390/cancers14225480 Agustina H, Asyifa I, Aziz A, Hernowo BS (2018) The role of os-
- teocalcin and alkaline phosphatase immunohistochemistry in os-teosarcoma diagnosis. Patholog. Res. Int. 2018: 6346409 https://doi. org/10.1155/2018/6346409
- Anderson WJ, Jo VY (2021) Diagnostic immunohistochemistry of soft tissue and bone tumors: An update on biomarkers that correlate with molecular alterations. Diagnostics 11: 690 https://doi. org/10.3390/diagnostics11040690
- Astolfi A, Fiore M, Melchionda F, Indio V, Bertuccio SN, Pession A (2019) BCOR involvement in cancer. Epigenomics 11: 835-855. https://doi.org/10.2217/epi-2018-0195 de Azevedo JWV, de Medeiros Fernandes TAA, Fernandes JV, de
- Azevedo JWV, de Medeiros Fernandes TAA, Fernandes JV, de Azevedo JCV, Lanza DCF, Bezerra CM, Andrade VS, de Araújo JMG, Fernandes JV (2020) Biology and pathogenesis of human osteosarcoma (Review). *Oncol. Lett.* **19**: 1099–1116. https://doi. org/10.3892/ol.2019.11229
- Brown LC, Tucker MD, Sedhom R, Schwartz EB, Zhu J, Kao C, Lab-riola MK, Gupta RT, Marin D, Wu Y, Gupta S, Zhang T, Harrison MR, George DJ, Alva A, Antonarakis ES, Armstrong AJ (2021) LRP1B mutations are associated with favorable outcomes to

immune checkpoint inhibitors across multiple cancer types. J. Immunother. Cancer 9: 1-10. https://doi.org/10.1136/jitc-2020-001

- Calvanese V, Nguyen AT, Bolan TJ, Vavilina A, Su T, Lee LK, Wang Y, Lay FD, Magnusson M, Crooks GM, Kurdistani SK, Mikkola HKA (2019) MLLT3 governs human haematopoietic stem-cell self-renewal and engraftment. Nature 576: 281-286. https://doi. org/10.1038/s41586-019-1790-2
- Chen Z, Guo J, Zhang K, Guo Y (2016) TP53 Mutations and survival in osteosarcoma patients: a meta-analysis of published data. Dis. Markers 2016 https://doi.org/10.1155/2016/4639575
- Cheng WL, Feng PH, Lee KY, Chen KY, Sun WL, Van Hiep N, Luo CS, Wu SM (2021) The role of EREG/EGFR pathway in tumor progression. Int. J. Mol. Sci. 22: 12828. https://doi.org/10.3390/ ijms222312828
- Chiappetta C, Carletti R, Della Rocca C, Di Cristofano C (2019) KMT2C modulates migration and invasion processes in osteosarcoma cell lines. Pathol. Res. Pract. 215: 152534. https://doi. org/10.1016/j.prp.2019.12534 Croce S, Chibon F (2015) MED12 and uterine smooth muscle onco-
- genesis: State of the art and perspectives. Eur. J. Cancer 51: 1603-
- Geness, state of the art and perspectives. Eur. J. Canter 31: 1005– 1610. https://doi.org/10.1016/j.ejca.2015.04.023
 Czarnecka AM, Synoradzki K, Firlej W, Bartnik E, Sobczuk P, Fie-dorowicz M, Grieb P, Rutkowski P (2020) Molecular biology of osteosarcoma. Cancers (Basel) 12: 1–27. https://doi.org/10.3390/cancers12082130
- Ding N, Zhou H, Esteve PO, Chin HG, Kim S, Xu X, Joseph SM, Friez MJ, Schwartz CE, Pradhan S, Boyer TG (2008) Mediator links epigenetic silencing of neuronal gene expression with X-linked mental retardation. Mol. Cell 31: 347-359. https://doi.org/10.1016/j. molcel.2008.05.023
- Fletcher CDM, Unni KK (2002) World Health Organization Classification of Tumours Pathology and Genetics of Tumours of Soft Tissue and Bone
- Gala K, Li Q, Sinha A, Razavi P, Dorso M, Sanchez-Vega F, Chung YR, Hendrickson R, Hsieh JJ, Berger M, Schultz N, Pastore A, Abdel-Wahab O, Chandarlapaty S (2018) KMT2C mediates the estrogen dependence of breast cancer through regulation of ERa enhancer function. Oncogene 37: 4692-4710. https://doi.org/10.1038/ 41388-018-0273-5
- Haase S, Garcia-Fabiani MB, Carney S, Altshuler D, Núñez FJ, Méndez FM, Núñez F, Lowenstein PR, Castro MG (2018) Mutant ATRX: uncovering a new therapeutic target for glioma. Expert Opin. Ther. Targets 22: 599-613. https://doi.org/10.1080/14728222.2018.1
- He Q, Kim H, Huang R, Lu W, Tang M, Shi F, Yang D, Zhang X, Huang J, Liu D, Songyang Z (2015) The Daxx/Atrx complex protects tandem repetitive elements during DNA hypomethylation by promoting H3K9 trimethylation. Cell Stem Cell 17: 273-286. https:/ doi.org/10.1016/j.stem.2015.07.022 He Z, Yang C, He Y, Gong B, Yin C, Feng J, Chen L, Tang J, Chen
- Y (2021) CAMTA1, a novel antitumor gene, regulates proliferation and the cell cycle in glioma by inhibiting AKT phosphorylation. *Cell. Signal.* **79**: 109882. https://doi.org/https://doi.org/10.1016/j. cellsig 2020 10988
- Huang Q, Li F, Hu H, Fang Z, Gao Z, Xia G, Ng WL, Khodadadi-Jamayran A, Chen T, Deng J, Zhang H, Almonte C, Labbe K, Han H, Geng K, Tang S, Freeman GJ, Li Y, Chen H, Wong KK (2022) Loss of TSC1/TSC2 sensitizes immune checkpoint block-ade in non-small cell lung cancer. *Sci. Adv.* **8**: 1–11. https://doi. org/10.1126/sciadv.abi9533
- Huang T, Zhou Y, Cao Y, Tao J, Zhou ZH, Hang DH (2017) STK39, overexpressed in osteosarcoma, regulates osteosarcoma cell in-vasion and proliferation. Oncol. Lett. 14: 4599-4604. https://doi.
- org/10.3892/ol.2017.6728 Huynh KD, Fischle W, Verdin E, Bardwell VJ (2000) BCoR, a novel corepressor involved in BCL-6 repression. *Genes Dev* 14: 1810–1823. Inoki K, Li Y, Zhu T, Wu J, Guan KL (2002) TSC2 is phosphoryl-
- ated and inhibited by Akt and suppresses mTOR signalling. Nat. Cell Biol. 4: 648–657. https://doi.org/10.1038/ncb839 Jelinic P, Mueller JJ, Olvera N, Dao F, Scott SN, Shah R, Gao J, Schultz N, Gonen M, Soslow RA, Berger MF, Levine DA (2014) Proverse SMARCA. Recurrent SMARCA4 mutations in small cell carcinoma of the ova-
- ry. Nat. Genet. 46: 424–426. https://doi.org/10.1038/ng.2922 Jones S, Wang T-L, Shih I-M, Mao T-L, Nakayama K, Roden R, Glas R, Slamon D, Diaz LAJ, Vogelstein B, Kinzler KW, Velculescu VE, Papadopoulos N (2010) Frequent mutations of chromatin remodeling gene ARID1A in ovarian clear cell carcinoma. Science 330: 228–231. https://doi.org/10.1126/science.1196333 Koelsche C, Schrimpf D, Tharun L, Roth E, Sturm D, Jones DTW,
- Renker E-K, Sill M, Baude A, Sahm F, Capper D, Bewerunge-Hudler M, Hartmann W, Kulozik AE, Petersen I, Flucke U, Schreuder HWB, Büttner R, Weber M-A, Schirmacher P et al. (2017) Histone 3.3 hotspot mutations in conventional osteosarcomas: a comprehensive clinical and molecular characterization of six H3F3A mutated cases. Clin. Sarcoma Res. 7: 9 https://doi. org/10.1186/s13569-017-0075-5

- Kuroda K, Moriya K, Nakano T, Saito R, Sato D, Katayama S, Niizuma H, Watanuki M, Uematsu M, Sasahara Y, Kure S (2021) A pediatric case of osteosarcoma and tuberous sclerosis complex with a novel germline mutation in the TSC2 gene and a somatic mutation in the TP53 gene. *Pediatr. Blood Cancer* 68: 2–3. https://doi. org/10.1002/pbc.28960
- Le MK, Omori M, Oishi N, Oi M, Fukasawa H, Hirata S, Kondo T (2021) High-grade uterine sarcoma with osteosarcomatous differentiation arising from a MED12-mutated leiomyoma, a case report. *Pathol. Int.* **71**: 199–203. https://doi.org/10.1111/pin.13065
- Lehnart SE, Wehrens XHT, Reiken S, Warrier S, Belevych AE, Harvey RD, Richter W, Jin SLC, Conti M, Marks AR (2005) Phosphodiesterase 4D deficiency in the ryanodine-receptor complex promotes heart failure and arrhythmias. *Cell* **123**: 25–35. https://doi. org/10.1016/j.cell.2005.07.030
- Lerwill MF, Lee AHS, Tan PH (2022) Fibroepithelial tumours of the breast-a review. Virchows Arch. 480: 45–63. https://doi.org/10.1007/ s00428-021-03175-6
- Li Y, Yang S, Liu Y, Yang S (2022) Deletion of Trp53 and Rb1 in Ctsk-expressing cells drives osteosarcoma progression by activating glucose metabolism and YAP signaling. *Med. Commun.* 3: 1–15. https://doi.org/10.1002/mco2.131
- Lian J, Xu C, Chen X, Huang S, Wu D (2022) Histone methyltransferase KMT2C plays an oncogenic role in prostate cancer. J. Cancer Res. Clin. Oncol. 148: 1627–1640. https://doi.org/10.1007/s00432-022-03968-5
- Limmahakhun S, Pothacharoen P, Theera-Umpon N, Arpornchayanon O, Leerapun T, Luevitoonvechkij S, Pruksakorn D (2011) Relationships between serum biomarker levels and clinical presentation of human osteosarcomas. *Asian Pacific J. Cancer Prev.* 12: 1717–1722
- human osteosarcomas. Asian Pacific J. Cancer Prev. 12: 1717–1722 Lin A, Qiu Z, Zhang J, Luo P (2021) Effect of NCOR1 mutations on immune microenvironment and efficacy of immune checkpoint inhibitors in patient with bladder cancer. Front. Immunol. 12: 630773. https://doi.org/10.3389/fimmu.2021.630773
- Liu X, Qiu R, Xu M, Meng M, Zhao S, Ji J, Yang Y (2021) KMT2C is a potential biomarker of prognosis and chemotherapy sensitivity in breast cancer. *Breast Cancer Res. Treat.* 189: 347–361. https://doi. org/10.1007/s10549-021-06325-1
- Lu P, Gu Y, Li L, Wang F, Yang X, Yang Y (2018) Long Noncoding RNA CAMTA1 Promotes proliferation and mobility of the human breast cancer cell line MDA-MB-231 via targeting miR-20b. Oncol. Res. 26: 625–635. https://doi.org/10.3727/09650401 7X14953948675395
- Luo Z, Xiao L, Li J, Dong B, Wang C (2019) Integrative analysis reveals driver long non-coding RNAs in osteosarcoma. *Medicine (Baltimore)* 98: e14302. https://doi.org/10.1097/MD.000000000014302
- Machado I, Navarro S, Picci P, Llombart-Bosch A (2016) The utility of SATB2 immunohistochemical expression in distinguishing between osteosarcomas and their malignant bone tumor minickers, such as Ewing sarcomas and chondrosarcomas. *Pathol. Res. Pract.* 212: 811– 816. https://doi.org/10.1016/j.prp.2016.06.012
- Marinö-Enriquez A, Lauria A, Przybyl J, Ng TL, Kowalewska M, Debiec-Rychter M, Ganesan R, Sumathi V, George S, Glenn McCluggage W, Nucci MR, Lee CH, Fletcher JA (2018) BCOR internal tandem duplication in high-grade uterine sarcomas. *Am. J. Surg. Pathol.* 42: 335–341. https://doi.org/10.1097/PAS.000000000000993
- Masliah-Planchon J, Lévy D, Héron D, Giuliano F, Badens C, Fréneaux P, Galmiche L, Guinebretierre JM, Cellier C, Waterfall JJ, Aït-Raïs K, Pierron G, Glorion C, Desguerre I, Soler C, Deville A, Delattre O, Michon J, Bourdeaut F (2018) Does ATRX germline variation predispose to osteosarcoma? Three additional cases of osteosarcoma in two ATR-X syndrome patients. *Eur. J. Hum. Genet.* 26: 1217–1221. https://doi.org/10.1038/s41431-018-0147-x
- Misaghi A, Goldin A, Awad M, Kulidjian AA (2018) Osteosarcoma : a comprehensive review. SICOT J. 4 https://doi.org/doi: 10.1051/ sicotj/2017028.
- Noblejas-López M del M, Morcillo-García S, Nieto-Jiménez C, Nuncia-Cantarero M, Győrffy B, Galan-Moya EM, Pandiella A, Ocaña A (2018) Evaluation of transcriptionally regulated genes identifies NCOR1 in hormone receptor negative breast tumors and lung adenocarcinomas as a potential tumor suppressor gene. *PLoS One* 13: 1–12. https://doi.org/10.1371/journal.pone.0207776
 Nomura R, Saito T, Mitomi H, Hidaka Y, Lee S, Watanabe S, Yao T
- Nomura R, Saito T, Mitomi H, Hidaka Y, Lee S, Watanabe S, Yao T (2014) GNAS mutation as an alternative mechanism of activation of the Wnt/β-catenin signaling pathway in gastric adenocarcinoma of the fundic gland type. *Hum. Pathol.* **45**: 2488–2496. https://doi. org/10.1016/j.humpath.2014.08.016
- Pan R, Zhang Z, Jia H, Ma J, Wu C, Xue P, Cai W, Zhang X, Sun J (2022) CAMTA1–PPP3CA–NFATc4 multi-protein complex mediates the resistance of colorectal cancer to oxaliplatin. *Cell Death Discon.* 8 https://doi.org/10.1038/s41420-022-00912-x
- Park S-M, Choi E-Y, Bae M, Kim S, Park JB, Yoo H, Choi JK, Kim Y-J, Lee S-H, Kim I-H (2016) Histone variant H3F3A promotes lung cancer cell migration through intronic regulation. *Nat. Commun.* 7 https://doi.org/10.1038/ncomms12914

- Pasche B (2001) Role of transforming growth factor beta in cancer. J. Cell. Physiol. 186: 153–168. https://doi.org/10.1002/1097-4652(200002)186:2<153::AID-JCP1016>3.0.CO;2-J
- Patra KC, Kato Y, Mizukami Y, Widholz S, Revenco I, Grossman EA, Ji F, Sadreyev RI, Liss AS, Screaton RA, Sakamoto K, Ryan DP, Mino- M, Castillo CF, Nomura DK, Haas W, Bardeesy N (2018) Mutant GNAS drives pancreatic tumourigenesis by inducing PKAmediated SIK suppression and reprogramming lipid metabolism. *Nat. Cell Biol.* 20: 811–822. https://doi.org/10.1038/s41556-018-0122-3.
- Piek E, Heldin CH, Ten Dijke P (1999) Specificity, diversity, and regulation in TGF-beta superfamily signaling. FASEB J 13: 2105–2124. PMID: 10593858
- Pignochino Y, Grignani G, Cavalloni G, Motta M, Tapparo M, Bruno S, Bottos A, Gammaitoni L, Migliardi G, Camussi G, Alberghini M, Torchio B, Ferrari S, Bussolino F, Fagioli F, Picci P, Aglietta M (2009) Sorafenib blocks tumour growth, angiogenesis and metastatic potential in preclinical models of osteosarcoma through a mechanism potentially involving the inhibition of ERK1/2, MCL-1 and ezrin pathways. *Mol. Cancer* 8: 1–17. https://doi.org/10.1186/1476-4598-8-118
- Porter, DE; Holden, ST; Steel, CM; Cohen, BB; Wallace, MR; Reid R (1992) A significant proportion of patients with osteosarcoma may belong to Li-Fraumeni cancer families. J. Bone Jt. Surg. 74: 883–886. https://doi.org/10.1302/0301-620x.74b6.1447251
- Príncipe C, Dionísio de Sousa IJ, Prazeres H, Soares P, Lima RT (2021) LRP1B: A Giant Lost in Cancer Translation. *Pharmaceuticals* (Basel) 14: https://doi.org/10.3390/ph14090836
- Sbaraglia M, Righi A, Gambarotti M, Dei Tos AP (2020) Ewing sarcoma and Ewing-like tumors. Virchows Arch. 476: 109–119. https:// doi.org/10.1007/s00428-019-02720-8
- Sharma S, Kelly TK, Jones PA (2009) Epigenetics in cancer. Carcinogenesis 31: 27–36. https://doi.org/10.1093/carcin/bgp220
 Sigismund S, Avanzato D, Lanzetti L (2018) Emerging func-
- Sigismund S, Avanzato D, Lanzetti L (2018) Emerging functions of the EGFR in cancer. Mol. Oncol. 12: 3–20. https://doi. org/10.1002/1878-0261.12155
- Śmiech M, Leszczyński P, Kono H, Wardell C, Taniguchi H (2020) Emerging BRAF mutations in cancer progression and their possible effects on transcriptional networks. *Genes (Basel)* 11: https://doi. org/10.3390/genes11111342
- Soejima H, Kawamoto S, Akai J, Miyoshi O, Arai Y, Morohka T, Matsuo S, Niikawa N, Kimura A, Okubo K, Mukai T (2001) Isolation of novel heart-specific genes using the BodyMap database. *Genomics* 74: 115–120. https://doi.org/10.1006/geno.2001.6527
- Sturm D, Witt H, Hovestadt V, Khuong-Quang DA, Jones DT, Konermann C, Pfaff E, Tönjes M, Sill M, Bender S, Kool M, Zapatka M, Becker N, Zucknick M, Hielscher T, Liu XY, Fontebasso AM, Ryzhova M, Albrecht S, Jacob K, Wolter M, Ebinger M, Schuhmann MU, van Meter T, Frühwald MC, Hauch H, Pekrun A, Radlwimmer B, Niehues T, von Komorowski G, Dürken M, Kulozik AE, Madden J, Donson A, Foreman NK, Drissi R, Fouladi M, Scheurlen W, von Deimling A, Monoranu C, Roggendorf W, Herold-Mende C, Unterberg A, Kramm CM, Felsberg J, Hartmann C, Wiestler B, Wick W, Milde T, Witt O, Lindroth AM, Schwartzentruber J, Faury D, Fleming A, Zakrzewska M, Liberski PP, Zakrzewski K, Hauser P, Garami M, Klekner A, Bognar L, Morrissy S, Cavalli F, Taylor MD, van Sluis P, Koster J, Versteeg R, Volckmann R, Mikkelsen T, Aldape K, Reifenberger G, Collins VP, Majewski J, Korshunov A, Lichter P, Plass C, Jabado N, Pfister SM (2012) Hotspot mutations in H3F3A and IDH1 define distinct epigenetic and biological subgroups of glioblastoma. *Cancer Cell* 22: 425–437. https://doi. org/10.1016/j.ccr.2012.08.024
- Sugiura Y, Kanda H, Motoi N, Nomura K, Inamura K, Okada E, Matsumoto H, Shimoji T, Matsumoto S, Nakayama J, Takazawa Y, Ishikawa Y, Machinami R (2018) Osteosarcoma arising in fibrous dysplasia, confirmed by mutational analysis of GNAS gene. *Pathol. -Res. Pract.* 214: 318–324. https://doi.org/https://doi.org/10.1016/j. prp.2017.10.018
- Sun J, Ren P, Ye L, Li N, Wang D (2017) MILT3 promotes proliferation of osteosarcoma cells by regulating JNK signaling. Int. J. Clin. Exp. Pathol. 10: 9444–9451
- Sun Ý, Wang L, Li C, Gu R, Zang W, Song W, Xia P (2020) Construction of an integrated human osteosarcoma database, HOsDb, based on literature mining, microarray analysis, and database retrieval. *BMC Cancer* 20: 1–9. https://doi.org/10.1186/s12885-020-06719-2
- Synoradzki KJ, Bartnik E, Czarnecka AM, Fiedorowicz M, Firlej W, Brodziak A, Stasinska A, Rutkowski P, Grieb P (2021) Tp53 in biology and treatment of osteosarcoma. *Cancers (Basel)*. 13: 1–23. https://doi.org/10.3390/cancers13174284
- Tallegas M, Gomez-Brouchet A, Legrand M, Bouvier C, de Pinieux G (2022) Markers for bone sarcomas. In Bone Sarcomas and Bone Metastases – From Bench to Bedside. Heymann D eds., pp 543–577. Academic Press. https://doi.org/https://doi.org/10.1016/B978-0-12-821666-8.00017-7

- Tang L, Zhang L, Liu L, Dong L, Dong Y, Zhu W, Wang H (2020) NCOR1 may be a potential biomarker of a novel molecular subtype of prostate cancer. FEBS Open Bio 10: 2678-2686. https://doi. org/10.1002/2211-5463.13004
- Tate JG, Bamford S, Jubb HC, Sondka Z, Beare DM, Bindal N, Boutselakis H, Cole CG, Creatore C, Dawson E, Fish P, Harsha B, Hathaway C, Jupe SC, Kok CY, Noble K, Ponting L, Ramshaw CC, Rye CE, Speedy HE, Stefancsik R, Thompson SL, Wang S, Ward S, Campbell PJ, Forbes SA (2019) COSMIC: The Catalogue of Somatic Mutations in Cancer. Nucleic Acids Res. 47: D941-D947. https://doi.org/10.1093/nar/gky1015 Tokoyoda M, Adachi S, Ishida Y, Yamazaki K (2018) Osteosarcoma
- mimic in the breast: A recurrent malignant phyllodes tumour harbouring MED12 and hTERT mutations. Cytopathology 29: 383-385. https://doi.org/10.1111/cyt.12544
- Trovó-Marqui AB, Tajara EH (2006) Neurofibromin: A general outlook. Clin. Genet. 70: 1-13. https://doi.org/10.1111/j.1399-0004.2006.00639.x
- Ueno-Yokohata H, Okita H, Nakasato K, Akimoto S, Hata JI, Koshinaga T, Fukuzawa M, Kivokawa N (2015) Consistent in-frame internal tandem duplications of BCOR characterize clear cell sarcoma of the kidney. Nat. Genet. 47: 861-863. https://doi.org/10.1038/ ng.3338
- Wang K, Yamamoto H, Chin JR, Werb Z, Vu TH (2004) Epidermal growth factor receptor-deficient mice have delayed primary endochondral ossification because of defective osteoclast recruitment. Biol. Chem. 279: 53848-53856. https://doi.org/10.1074/jbc. M403114200
- Wang M, Xiong Z (2021) The mutation and expression level of Irp1b are associated with immune infiltration and prognosis in hepatocellular carcinoma. Int. J. Gen. Med. 14: 6343-6358. https://doi. org/10.2147/IJGM.S333390
- Wang SL, Zhong GX, Wang XW, Yu FQ, Weng DF, Wang XX, Lin JH (2018) Prognostic significance of the expression of HER family members in primary osteosarcoma. Oncol. Lett. 16: 2185–2194. https://doi.org/10.3892/ol.2018.8931 Won KY, Kim YW, Park YK (2010) Expression of Smad and its sig-
- nalling cascade in osteosarcoma. Pathology 42: 242-247. https://doi. org/10.3109/00313021003631288
- Wu CC, Livingston JA (2020) Genomics and the immune landscape of osteosarcoma. Adv. Exp. Med. Biol. 1258: 21-36. https://doi. org/10.1007/978-3-030-43085-6_2

- Wu F, Zhong Y, Lang XB, Tu YL, Sun SF (2019) MNX1-AS1 accelerates the epithelial-mesenchymal transition in osteosarcoma cells by activating MNX1 as a functional oncogene. Eur. Rev. Med. Pharmacol. Sci. 23: 8194-8202. https://doi.org/10.26355/eurrev_201910_19126
- Xie Y, Huang J, Wu M, Zhou Y (2018) Expression of CD133 protein in osteosarcoma and its relationship with the clinicopathological features and prognosis. J. Cancer Res. Ther. 14: 892–895. https://doi.org/https://doi.org/10.4103/jcrt.JCRT_461_17
 Xu L, Xie X, Shi X, Zhang P, Liu A, Wang J, Zhang B (2021) Potential application of genomic profiling for the diagnosis and treat-
- ment of patients with sarcoma. Oncol. Lett. 21: 1-12. https://doi. org/10.3892/ol.2021.12614
- Xu N, Wang L, Sun P, Xu S, Fu S, Sun Z (2019) Low Arid1a expression correlates with poor prognosis and promotes cell proliferation and metastasis in osteosarcoma. Pathol. Oncol. Res. 25: 875-881. https://doi.org/10.1007/s12253-017-0338-8
- Yan GN, Lv YF, Guo QN (2016) Advances in osteosarcoma stem cell research and opportunities for novel therapeutic targets. Cancer Lett. 370: 268-274. https://doi.org/10.1016/j.canlet.2015.11.003
- Yang C, Tian Y, Zhao F, Chen Z, Su P, Li Y, Qian A (2020) Bone microenvironment and osteosarcoma metastasis. Int. J. Mol. Sci. 21: 1-17. https://doi.org/10.3390/ijms21196985
- Yang G, Yuan J, Li K (2013) EMT transcription factors: Implication in osteosarcoma. Med. Oncol. 30: 1–5. https://doi.org/10.1007/s12032-013-0697-2
- Yiqi Z, Ziyun L, Qin F, Xingli W, Liyu Y (2020) Identification of 9-gene epithelial-mesenchymal transition related signature of osteosarcoma by integrating multi cohorts. Technol. Cancer Res. Treat. 19: 1-11. https://doi.org/10.1177/1533033820980769 Yu X, Yustein JT, Xu J (2021) Research models and mesenchymal/
- epithelial plasticity of osteosarcoma. *Cell Biosci.* **11**: 1–12. https://doi. org/10.1186/s13578-021-00600-w
- Zauber P, Marotta SP, Sabbath-Solitare M (2016) GNAS gene mutation may be present only transiently during colorectal tumorigenesis. Int. J. Mol. Epidemiol. Genet. 7: 24-31.
- Zhou Y, Tan Y, Zhang Q, Duan Q, Chen J (2022) MED12 mutation as a potential predictive biomarker for immune checkpoint inhibitors in pan-cancer. Eur. J. Med. Res. 27: 1-5. https://doi. org/10.1186/s40001-022-00856-z